General

From Bone To Blood Flow: Tissue Engineering In Orthopedics – A Narrative Review

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Musculoskeletal injuries and degenerative conditions necessitate advanced regenerative solutions. Tissue engineering has emerged as a pivotal field in orthopedic care, particularly in vascularized bone and cartilage regeneration. This narrative review examines the latest advancements in vascular tissue engineering, including scaffold design, cell-based techniques, and growth factor delivery. A comprehensive literature search was conducted using PubMed, ScienceDirect, and Google Scholar, focusing on innovations and challenges in the field. Vascularized bone grafts (VBGs) outperform non-vascularized counterparts in promoting healing and integration. Advances in scaffold materials, such as smart scaffolds and hybrid biomaterials, enhance osteogenesis and angiogenesis. Cellular therapies, utilizing mesenchymal stem cells and induced pluripotent stem cells, synergistically improve vascularization and bone regeneration. Growth factors like VEGF and bone morphogenic protein (BMP-2), integrated with innovative delivery systems, enable sustained angiogenic stimulation and scaffold integration. While significant strides have been made, challenges persist in achieving full vascular integration and replicating native tissue architecture. Innovations in scaffold technology and vascular surgery techniques hold promise for transforming orthopedic tissue engineering and improving patient outcomes.

I. INTRODUCTION

Musculoskeletal, bone, and cartilage injuries are increasing due to demographic aging and the growing focus on exercise, underscoring the need for advanced treatment and rehabilitation strategies to enhance patient care and quality of life. The emergence of regenerative medicine has paved the way for successfully recreating healthy tissues. 2

Regenerative medicine, a rapidly advancing field, encompasses tissue engineering as a critical component that seeks to restore damaged tissues and improve overall functionality. Recent innovations in orthopedic tissue engineering focus on regenerating bone, cartilage, and musculoskeletal tissues using engineered constructs to improve patient outcomes and quality of life.³ The trajectory of orthopedic surgery has shifted with the advent of minimally invasive surgical techniques that reduce surgical and postoperative trauma.⁴

Orthopedic tissue regeneration relies on four essential components: cells, signals, scaffolds, and a suitable mechanical environment. Strategies to combine these components are intensively researched to ensure clinically effective tissue regeneration.³

Grafts can be developed from various stem cell types, such as mesenchymal, embryonic, and pluripotent stem cells, each evaluated for their potential in tissue regeneration. Scaffolds, which can be derived from natural materials, synthetic substances, or devitalized extracellular matrices, offer benefits and limitations depending on their source.⁵

Given the highly grafted nature of bone, vascularization is crucial to ensure successful tissue integration and long-term graft viability. Vascular tissue engineering aims to create functional vascular networks within engineered tissues, facilitating their growth and integration into the host. Integrating vascular surgery techniques into orthopedic tis-

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sue engineering holds promise for addressing complex cases like non-union fractures and avascular necrosis. ^{7,8}

This review examines the advancements, challenges, and future directions of vascular tissue engineering in orthopedics, focusing on innovations in scaffold design, biomaterials, progenitor cell grafts, angiogenic factor delivery, and the application of vascularized grafts for bone, cartilage, and joint repair.

II. MATERIALS AND METHODS

A comprehensive literature review was conducted to explore vascularized bone grafts and tissue engineering approaches in orthopedics. The search included articles published in peer-reviewed journals, accessed through PubMed, ScienceDirect, and Google Scholar. Search terms included "vascularized bone grafts," "orthopedic tissue engineering," "scaffold design," "vascular tissue engineering," "angiogenesis," and "growth factor delivery." Studies were selected based on relevance to bone and cartilage regeneration, with an emphasis on recent advancements in scaffold materials, cell-based therapies, and vascularization techniques.

III. THE ROLE OF VASCULARIZATION STRATEGIES IN ENHANCING BONE AND CARTILAGE HEALING

IMPORTANCE OF VASCULARIZATION

Non-union fractures remain a frequent complication in orthopedic surgery despite significant research advancements. Adequate vascularization plays a key role in ensuring successful bone regeneration by providing the callus tissue with the essential oxygen, nutrients, and growth factors. Similarly, the successful treatment of large segmental bone defects is also hindered by factors like infection, tumor resection, and soft tissue damage. Thus, effective healing relies heavily on restoring a large volume of bone, making vascularization strategies crucial for supporting bone regeneration and remodeling. ¹⁰

VASCULARIZED BONE GRAFTS

Vascularized bone grafts (VBGs) have proved superior to non-vascularized bone grafts (NVBGs) because VBGs contain living osteocytes and active blood flow, allowing for direct healing and better preservation of biomechanical properties and mineral content, thereby reducing bone resorption and enhancing healing potential.¹¹ Unlike NVBGs, which rely on creeping substitution through a dead bone matrix, VBGs can effectively remodel and integrate into the recipient site even in avascular environments, preventing complications like non-unions, infections, and graft failure.^{12,13}

<u>Table 1</u> represents a detailed comparison of various vascularized bone grafts, outlining their locations, indications, vascular contributions, potential complications, and additional clinical considerations. ^{11,14-21} <u>Figure 1</u> represents

an overview of scaffold-based platforms for stem cell and growth factor delivery in bone tissue engineering.

VASCULAR SURGERY TECHNIQUES FOR LIMB SALVAGE

In addition to the use of vascularized bone grafts for reconstructing non-union fractures and large bone defects, vascular surgery techniques play an equally critical role in limb salvage procedures. In cases of severe trauma or compromised blood flow, techniques such as arterial bypass and vein grafting become essential for restoring circulation and ensuring the viability of both soft tissue and bone. Lower extremity bypass surgery involves using an autogenous vein, like the ipsilateral greater saphenous vein (GSV), to bypass arterial obstructions, identified through preoperative vascular imaging, from a proximal to a distal site.²² This procedure is especially crucial for patients with chronic limb-threatening ischemia and distal occlusive disease, where autologous veins are preferred over prosthetic grafts for better long-term outcomes.²³ The small saphenous vein (SSV) can also be used in patients when harvesting the GSV is difficult.²⁴

Advancing beyond current approaches, the integration of novel scaffold designs, improved growth factor delivery systems, and cell-based techniques are driving the future of vascular tissue engineering in orthopedics.

IV. ADVANCES IN VASCULAR TISSUE ENGINEERING FOR BONE REGENERATION

Although VBGs are commonly used for bone repair, drawbacks such as the limited availability of autologous bone grafts and issues like donor site morbidity, along with the potential for rejection in allogeneic grafts, can increase the risk of implant failure. ²⁵ Thus, engineered vascular tissues have emerged as a viable alternative, with an emphasis on incorporating pro-angiogenic factors like vascular endothelial growth factor (VEGF) into scaffolds to promote the blood vessel formation within the engineered constructs, facilitating osteogenesis and tissue integration. ²⁵ Additionally, scaffold designs that mimic the natural bone micro-environment can significantly improve the vascularization process and lead to faster bone regeneration. ²⁶

A. CELLULAR TECHNIQUES

Cellular techniques are pivotal in creating functional vascular networks for bone regeneration in orthopedic applications. Endothelial cells (ECs) initiate angiogenesis by forming the framework for vascular networks, ensuring nutrient delivery, oxygen supply, and waste removal critical for bone healing. Mesenchymal stem cells (MSCs) enhance vascularization by differentiating into multiple cell types and releasing pro-angiogenic factors like VEGF, supporting endothelial proliferation and maturation. Induced pluripotent stem cells (iPSCs) provide scalability and potential for patient-specific therapy, crucial for long-term vascular integration. Advances in nano-polypeptide hy-

Table 1. Comparison of vascularized bone grafts for locations, indications and clinical considerations

Graft Location	Bone Type and Description	Vascular Contribution	Indications	Morbidity/ Complications
Fibula	Tubular bone, segment length 26-30 cm	Peroneal artery	Large segmental bone defects, mandibular reconstruction (esp. double-barrel technique for anterior defects), femoral head necrosis	Donor-site morbidity (e.g., peroneal nerve injury, lower extremity blood flow)
Medial Femoral Condyle	Cortico-cancellous, cortical surfaces, and potential osteo-cartilaginous structure	Descending genicular artery	Small to moderate bone defects, scaphoid non-union, avascular necrosis of carpal bones, extremity defects	Temporary knee pain common post-surgery
Distal Radius (Pedicled)	Cortical bone, limited volume, vascularized pedicled graft for small reconstructions	Radial artery	Small bone defects, particularly in carpal bones (e.g., scaphoid non- union, avascular necrosis of scaphoid)	Limited bone tissue, risk of donor site fracture, scarring
Iliac Crest	Cortico-cancellous, large concave shape	Deep circumflex iliac artery (DCIA)	Structural and contour defects in mandible, maxillofacial, and upper/lower extremity reconstructions	Risk of contour deformity, infection, chronic pain
Scapula	Multilamellar structure, cortical and cancellous	Angular branch of thoracodorsal artery (TDa)	Moderate to large bone defects, complex composite tissue defects, especially in elderly patients with vascular disease	Shoulder and arm dysfunction, seroma
Rib (Costal)	Membranous bone with dual blood supply	Posterior intercostal artery and periosteal supply from serratus anterior (via thoracodorsal artery)	Lower/upper extremity (e.g., humerus, clavicle, carpal non- union) and maxillofacial defects, jawbone substitute for mandible	Infection, chronic pain
Clavicle	Cortico-cancellous, 3-7 cm	Supraclavicular artery, superior thyroid artery	Shoulder and chest wall defects	Reduced shoulder strength, donor- site morbidity
Metatarsal (second)	Cortico-cancellous	Descending genicular artery	Small defect reconstruction in the foot	Pain, discomfort

drogels and 3D printing improve scaffold designs, facilitating stem cell differentiation and angiogenesis.³⁰

The integration of cell-based approaches in orthopedic tissue engineering represents a promising avenue for developing vascularized bone constructs. Across multiple studies, the synergistic use of MSCs and ECs has emerged as a fundamental strategy to promote both angiogenesis and osteogenesis, enhancing bone healing in large defects.^{29,} 31-33

The inclusion of induced pluripotent stem cells (iPSCs) adds versatility, allowing for differentiation into multiple cell types and enabling patient-specific therapies, which are critical for achieving better integration and functional outcomes in engineered bone tissues. ^{34,35} Replicating natural developmental processes, such as endochondral ossification, further strengthens the formation of mature bone structures, underscoring the importance of physiological pathways in engineering vascularized bone. ^{32,36}

Innovations in scaffold-free techniques, like temperature-responsive cell sheets, allow for the autonomous formation of functional tissue analogues.³³ These methods not only enhance scalability but also improve vascular in-

tegration, addressing critical challenges in treating complex orthopedic defects. Turther innovations in cell-based approaches include automated systems that streamline the production of multi-layered tissue constructs, enhancing scalability and reproducibility. These systems utilize robotic platforms to assemble cell sheets, enabling efficient production of pre-vascularized structures suitable for large, complex orthopedic defects. The Weever, given their recent advent, studies are yet to yield decisive data pertaining to the advantages of using such systems.

B. GROWTH FACTOR DELIVERY

Effective growth factor delivery is essential for promoting angiogenesis and bone regeneration in orthopedic tissue engineering. VEGF drives blood vessel formation, while factors like basic fibroblast growth factor (bFGF), platelet-derived growth factor (PDGF), and angiopoietins stabilize and mature vascular networks, crucial for integrating engineered constructs into bone tissues.³⁸ Strategies such as incorporating VEGF into biodegradable scaffolds provide sustained angiogenic stimulation during scaffold degrada-

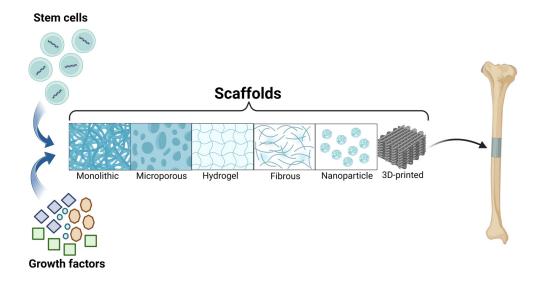


Figure 1. Overview of scaffold types in bone tissue engineering

tion, fostering vascular growth and bone healing.³⁹ Combining VEGF with bFGF enhances endothelial cell proliferation and migration, accelerating early vessel formation.⁴⁰, ⁴¹ Nanoparticle encapsulation of VEGF and PDGF enables controlled release, stabilizing vascular networks within bone defects.⁴²

Hydrogels, with their hydrophilic properties, deliver VEGF and angiopoietin-1 locally and efficiently, ensuring mature vascular networks for enhanced osteogenesis. 43 Gene delivery techniques further enable sustained VEGF production, supporting long-term angiogenesis in large bone defects, particularly when combined with bFGF or PDGF for vascular stabilization. 44,45

Electrospun fibers and microspheres integrated into scaffolds offer sequential release of VEGF, bFGF, and PDGF, promoting vascular density and maturity essential for orthopedic repair. 46,47 These approaches ensure targeted angiogenesis, creating vascularized environments critical for effective bone regeneration.

Recent studies have explored various applications of factors like BMPs and PDGF in bone tissue engineering, focusing on effective delivery methods and combinatory approaches to enhance outcomes. For instance, PDGF-BB loaded onto tricalcium phosphate (TCP) particles is noted for promoting cell recruitment and revascularization, with applications in fractures and joint fusions. In clinical trials, it demonstrated comparable effectiveness to autografts, with over 66% fusion rates observed at 24 weeks postoperatively.

FGF2 and BMP-2, combined with VEGF on silica-coated nanohydroxyapatite and gelatin scaffolds, were shown to stimulate vascularization and bone regeneration in calvarial defects, with varied release rates across growth factors. ⁵¹ For BMP-2, polyelectrolyte film coatings on hollow

poly(lactic-co-glycolic) acid (PLGA) tubes allowed tunable release, facilitating rapid bone formation in femoral defects. 52 A similar focus on BMP-2 and TGF-beta1 was seen in silk fibroin-based scaffolds, which promoted mesenchymal and osteoblast proliferation, enhancing osseointegration. 53

The combination of BMP-2 and FGF-2 on gelatin nanofibers stimulated osteogenic gene expression, while PDGF on poly-lactic acid (PLLA) nanofibers promoted vascularized bone regeneration in mouse calvarial defects. ⁵⁴⁻⁵⁶ Finally, VEGF and BMP-2 incorporated into microspheres with chitosan hydrogels achieved sustained release, leading to bone repair and callus remodeling in rabbit mandibular defects over 12 weeks. ⁵⁶ Table 2 represents a summary of key growth factors utilized in bone tissue engineering, highlighting their sources, biological roles, mechanisms of action, and primary functions when integrated into scaffold-based applications. ⁵⁷⁻⁶²

C. SCAFFOLD-BASED APPROACHES

Scaffold-based techniques are central to vascular tissue engineering, designed to replicate the extracellular matrix (ECM) and support cell growth, differentiation, and vessel formation. These scaffolds use a variety of biomaterials—natural, synthetic, or a combination of both—tailored to mimic the structural and functional properties of native tissues.

Natural polymers such as collagen, gelatin, and fibrin are biocompatible and degrade into non-toxic by-products, promoting endothelial cell proliferation, ECM synthesis, and osteoblast adhesion and integration.⁶⁴ In contrast, synthetic options like polylactic acid (PLA), polycaprolactone (PCL), and polyglycolic acid (PGA) provide superior

Table 2. Growth factors utilized in bone tissue engineering through direct application or scaffold delivery

Growth Factor	Primary Source	Key Biological Roles	Mechanism of Action	Main Functions
Fibroblast Growth Factor	Mesenchymal, Osteoblast, Chondrocyte, Inflammatory Cells, Endothelial	Promotes angiogenesis and tissue growth	Angiogenesis, proliferation, osteogenic differentiation	Encourages osteoblast proliferation; potent in wound healing and tissue repair
Vascular Endothelial Growth Factor	Platelets, Osteoblast, Chondrocyte	Supports angiogenesis and chemotactic signaling	Angiogenesis	Key in endothelial cell proliferation and survival, supplying nutrients through new blood vessels
Bone Morphogenetic Protein	Mesenchymal, Osteoblast, Endothelial, Chondrocyte	Drives bone formation, chondrogenic and osteogenic	Bone induction	Stimulates MSC migration and differentiation into osteoblasts, promoting bone formation without activating osteoclasts
Insulin-Like Growth Factor	Osteoblast, Chondrocyte, Hepatocyte, Endothelial	Anabolic effects on bone matrix synthesis	Osteogenic stimulation	Enhances osteoblast proliferation, bone matrix production, and stimulates osteoclasts
Platelet- Derived Growth Factor	Platelets, Osteoblast, Inflammatory Cells, Endothelial	Facilitates cell proliferation and tissue repair	Cell proliferation, vascularization	Central to wound healing and bone repair by promoting cell growth and angiogenesis
Transforming Growth Factor- β	Platelets, Osteoblast, Chondrocyte, Inflammatory Cells, Fibroblast, Endothelial	Osteoinductive, modulates immune responses	Differentiation of chondrogenic and osteogenic pathways	Stimulates bone growth by enhancing osteoprogenitor cells, ECM synthesis, and controlling osteoclast activity
Hepatocyte Growth Factor	Fibroblast, Endothelial Cells	Stimulates cell motility and proliferation	Mitogenic and morphogenic signaling	Promotes angiogenesis and tissue regeneration, often aiding in liver and wound repair processes
Epidermal Growth Factor	Platelets, Salivary Glands	Enhances epithelial and fibroblast growth	Cell proliferation, epithelialization	Accelerates wound healing and skin regeneration, important for epithelial cell growth and repair

control over mechanical properties, degradation rates, and resistance to physiological stresses, 65,66 making them ideal for load-bearing orthopedic applications. Hybrid scaffolds, combining natural and synthetic materials, enhance osteoconductivity while maintaining structural integrity, enabling vessel maturation, bone formation, and defect healing. 67,68

Advanced fabrication techniques, including 3D printing and electrospinning, allow precise control over scaffold porosity, pore size, and distribution. These features are critical for nutrient diffusion, capillary network formation, and cellular infiltration, directly supporting vascular and bone regeneration. Furthermore, modifications to scaffold surface roughness (0.2–2 μm) and charge properties optimize osteoblast activity while preventing bacterial adhesion, reducing the risk of infection in orthopedic implants. 70,71

Smart Scaffolds represent a significant innovation in orthopedic tissue engineering, enabling scaffolds to sense and respond to external and internal stimuli.⁷² These include exogenous triggers, such as light or mechanical forces, and endogenous factors, like inflammation or bacterial activity, which can activate localized drug delivery

or adaptive scaffold deployment.⁷² Such dynamic systems align with the functional demands of bone repair and regeneration, making smart scaffolds highly effective for targeted therapeutic outcomes.

The impact of a scaffold on native cells is a key factor in material selection. Scaffolds can be osteoconductive, allowing bone growth (like calcium sulfates), or osteoinductive, actively stimulating bone formation in defects that would not heal on their own (like demineralized bone matrix). Recent advancements in scaffold design for orthopedics focus on enhancing vascularization, bone regeneration, and infection prevention. Homework Biomaterials like modified collagen, elastin-like polymers, and bioactive ceramics improve cell adhesion, proliferation, and differentiation while 3D-printed scaffolds allow precise control of porosity, promoting nutrient exchange and capillary growth. Figure highlights the key factors and technologies in scaffold design for bone regeneration.

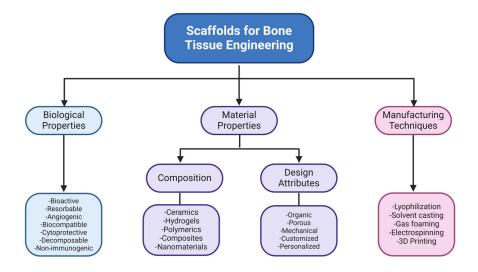


Figure 2. Overview of scaffold design for bone tissue engineering in terms of biological properties, material properties, and manufacturing techniques

V. TISSUE ENGINEERING CHALLENGES IN ORTHOPEDICS

Despite current advancements in vascular tissue engineering, there remain unique limitations in generating vascular tissue that retains biological functionality, particularly in orthopedics. Vascularization of load-bearing tissues such as bone and cartilage require full graft integration on multiple hierarchical levels which proves to be challenging due to graft necrosis, limited diffusion of nutrients, and lack of integration with host issue.⁷⁷ Bone tissue is composed of extracellular matrix signaling factors and bone cells which function in the formation and remodeling of natural bone tissue, and any graft intended to repair and regenerate bone must contain osteoconductive, osteoinductive, and osteogenic properties.⁷⁸ Vasculature repair in orthopedic tissues requires large perfusable blood vessel grafts at the injury site, then microvascular beds are needed as bone tissue faces a diffusion limit of 100–300 µm. 78,79 To obtain functionality of grafted large blood vessels, specially designed biomaterials must be used to mimic the structural compositions of the different vascular layers. 80 Some major limitations include recreating the distribution of cells within the three layers of vascular tissues in their natural arrangements. Moreover, reproducing a biocompatible surface that contains anti-thrombogenic properties remains a challenge. 81 The gold standard for bone repair is comprised of autologous grafts which improve vascularization; however, the autograft supply is limited and can cause pain at the donor site.⁷⁸ Sourcing decellularized xenogenic tissues for grafts has been used, which can avoid some complications of autologous grafts; however, severe immune responses to the graft may arise. 82 Recent advances include the development of porous bioactive glass-ceramic scaffolds which enhance nutrient flow, yet the challenge of vascularization remains present. 83 Another proposed alternative is using soft materials such as hydrogels for matrices as they can sustain cell viability for long periods of time; however, they face difficulty in integrating within the host tissue as their structure is easy to disrupt. 57,84

Moreover, innervation has become a new focus in the development of vascularized tissue-engineered bone. Restoring sensory innervation and suppressing sympathetic nerve activation have demonstrated the ability to enhance the osteogenic properties of functional scaffolds.⁸⁵

Despite the myriad advancements in the field of reconstructive surgery, one chief hurdle that remains to be overcome is the intrinsic properties of cartilage in being avascular – as well as aneural and alymphatic. ⁸⁶ However, recent studies have manifested the ability to vascularize tissueengineered cartilage in animal models, which may pave the way for important clinical implications. ^{87,88}

Overall, vascularization in tissue engineering faces challenges such as incomplete vascular integration, limited diffusion of nutrients, long-term viability, difficulty in replicating native vascular architectures as well as obstacles in immune response and the mechanical stability of scaffolds.

VI. CONCLUSION

Orthopedic tissue engineering, especially in the realm of vascularized bone and cartilage regeneration, holds immense potential for addressing complex musculoskeletal injuries and degenerative conditions. Significant advancements in scaffold design, cell-based therapies, and growth

factor delivery have demonstrated promise in promoting vascularization, essential for successful tissue integration and long-term graft viability. However, formidable challenges remain, particularly in achieving full vascular integration and replicating the hierarchical structure of native bone and cartilage. Innovations such as smart scaffolds, hybrid biomaterials, and gene delivery systems are paving the way toward more effective therapeutic options. Future research should prioritize overcoming the intrinsic limitations in scaffold vascularization and optimizing graft mechanical stability to improve clinical outcomes. The integration of vascular and orthopedic engineering techniques may ultimately establish new standards of care, enhancing patient recovery and quality of life.

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AUTHORS' CONTRIBUTIONS

BS, VG, KE, and MH contributed towards the concept of the study, the literature review, data extraction, and writing the different sections of the manuscript. BS worked on <u>Figure 1</u> and <u>table 2</u>. VG worked on <u>table 1</u>. MH participated in

data extraction, writing the manuscript, its rigorous revision several times, and coordinated the flow of the work. MN is the corresponding author. He made substantial contributions to the concept of the study and followed up the work to ensure that it was original and complied with the research ethics and guidelines of this journal. MN also contributed to the development of the manuscript. All authors are accountable for all the contributions and integrity of the work. All authors read and approved the final manuscript.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

Ethical review was not required because the study is a review of literature.

COMPETING INTERESTS

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REFERENCES

- 1. Cheng R, Wang H, Cheng CK. Advanced Engineering Technology in Orthopedic Research. *Bioengineering*. 2024;11(9):925. doi:10.3390/bioengineering11090925. PMID:39329667
- 2. Esdaille CJ, Washington KS, Laurencin CT. Regenerative engineering: a review of recent advances and future directions. *Regen Med*. 2021;16(5):495-512. doi:10.2217/rme-2021-0016
- 3. Alonzo M, Primo FA, Kumar SA, et al. Bone tissue engineering techniques, advances, and scaffolds for treatment of bone defects. *Current opinion in biomedical engineering*. 2021;17:100248. doi:10.1016/j.cobme.2020.100248. PMID:33718692
- 4. Liang W, Zhou C, Bai J, et al. Current advancements in therapeutic approaches in orthopedic surgery: a review of recent trends. *Frontiers in Bioengineering and Biotechnology*. Published online February 9, 2024:12. doi:10.3389/fbioe.2024.1328997
- 5. Bianchi E, Vigani B, Viseras C, Ferrari F, Rossi S, Sandri G. Inorganic Nanomaterials in Tissue Engineering. *Pharmaceutics*. 2022;14(6):1127. doi:10.3390/pharmaceutics14061127. PMID:35745700
- 6. Liu Y, Yang S, Cao L, Zhang X, Wang J, Liu C. Facilitated vascularization and enhanced bone regeneration by manipulation hierarchical pore structure of scaffolds. *Mater Sci Eng C Mater Biol Appl.* 2020;110:110622. doi:10.1016/j.msec.2019.110622
- 7. Chang WG, Niklason LE. A short discourse on vascular tissue engineering. *npj Regenerative Medicine*. 2017;2(1):7. doi:10.1038/s41536-017-0011-6
- 8. Shakeel A, Corridon PR. Mitigating challenges and expanding the future of vascular tissue engineering—are we there yet? Opinion. *Frontiers in Physiology*. Published online January 4, 2023:13. doi:10.3389/fphys.2022.1079421
- 9. Menger MM, Laschke MW, Nussler AK, Menger MD, Histing T. The vascularization paradox of non-union formation. *Angiogenesis*. 2022;25(3):279-290. doi:10.1007/s10456-022-09832-x
- 10. Schulze F, Lang A, Schoon J, Wassilew GI, Reichert J. Scaffold Guided Bone Regeneration for the Treatment of Large Segmental Defects in Long Bones. *Biomedicines*. 2023;11(2):325. doi:10.3390/biomedicines11020325. PMID:36830862

- 11. Archual AJ, Bishop AT, Shin AY. Vascularized Bone Grafts in Orthopaedic Surgery: A Review of Options and Indications. *JAAOS Journal of the American Academy of Orthopaedic Surgeons*. 2022;30(2):60-69. doi:10.5435/jaaos-d-20-01200
- 12. Testa G, Lucenti L, D'Amato S, et al. Comparison between Vascular and Non-Vascular Bone Grafting in Scaphoid Nonunion: A Systematic Review. *Journal of Clinical Medicine*. 2022;11(12):3402. doi:10.3390/jcm11123402. PMID:35743472
- 13. Feltri P, Solaro L, Errani C, Schiavon G, Candrian C, Filardo G. Vascularized fibular grafts for the treatment of long bone defects: pros and cons. A systematic review and meta-analysis. *Arch Orthop Trauma Surg.* 2023;143(1):29-48. doi:10.1007/s00402-021-03962-5
- 14. Abdulwadood I, Gomez DA, Martinez C, et al. Vascularized bone grafts in spinal reconstruction: An updated comprehensive review. *Orthoplastic Surgery*. 2024;17:7-14. doi:10.1016/j.orthop.2024.08.003
- 15. Holnthoner W, Banfi A, Kirkpatrick J, Redl H. *Vascularization for Tissue Engineering and Regenerative Medicine*. Springer; 2021. doi:10.1007/978-3-319-54586-8
- 16. Lonie S, Herle P, Paddle A, Pradhan N, Birch T, Shayan R. Mandibular reconstruction: meta-analysis of iliac-versus fibula-free flaps. *ANZ journal of surgery*. 2016;86(5):337-342. doi:10.1111/ans.13274
- 17. Pietruski P, Majak M, Światek-Najwer E, et al. Navigation-guided fibula free flap for mandibular reconstruction: A proof of concept study. *Journal of Plastic, Reconstructive & Aesthetic Surgery*. 2019;72(4):572-580. doi:10.1016/j.bjps.2019.01.026
- 18. Jones Jr D, Shin A. Medial femoral condyle vascularized bone grafts for scaphoid nonunions. *Chirurgie de la main*. 2010;29:S93-S103. doi:10.1016/j.main.2010.09.009
- 19. Wang W, Yeung KWK. Bone grafts and biomaterials substitutes for bone defect repair: A review. *Bioact Mater*. 2017;2(4):224-247. doi:10.1016/j.bioactmat.2017.05.007
- 20. Derby BM, Murray PM, Shin AY, et al. Vascularized bone grafts for the treatment of carpal bone pathology. *Hand (N Y)*. 2013;8(1):27-40. doi:10.1007/s11552-012-9479-0

- 21. Karaismailoglu B, Fatih Guven M, Erenler M, Botanlioglu H. The use of pedicled vascularized bone grafts in the treatment of scaphoid nonunion: clinical results, graft options and indications. *EFORT Open Rev.* 2020;5(1):1-8. doi:10.1302/2058-5241.5.190021
- 22. El-Sayed HF. Bypass surgery for lower extremity limb salvage: vein bypass. *Methodist Debakey Cardiovasc J.* 2012;8(4):37-42. doi:10.14797/mdcj-8-4-37
- 23. Rocha H, Antunes I, Rego D, et al. In situ lesser saphenous vein bypass through a posterior approach: an underestimated approach for limb salvage. *Angiologia e Cirurgia Vascular*. 2021;17(3):274-277.
- 24. Nierlich P, Metzger P, Enzmann FK, et al. The Small Saphenous Vein: An Underestimated Source for Autologous Distal Vein Bypass. *European Journal of Vascular and Endovascular Surgery*. 2019;58(4):556-563. doi:10.1016/j.eivs.2019.01.023
- 25. Pennings I, van Dijk LA, van Huuksloot J, et al. Effect of donor variation on osteogenesis and vasculogenesis in hydrogel cocultures. *Journal of Tissue Engineering and Regenerative Medicine*. 2019;13(3):433-445. doi:10.1002/term.2807
- 26. Lv N, Zhou Z, Hou M, et al. Research progress of vascularization strategies of tissue-engineered bone. Review. *Frontiers in Bioengineering and Biotechnology*. Published online January 19, 2024:11. doi:10.3389/fbioe.2023.1291969
- 27. Lovett M, Lee K, Edwards A, Kaplan DL. Vascularization strategies for tissue engineering. *Tissue Eng Part B Rev.* 2009;15(3):353-370. doi:10.1089/ten.TEB.2009.0085
- 28. Tao H, Han Z, Han ZC, Li Z. Proangiogenic Features of Mesenchymal Stem Cells and Their Therapeutic Applications. *Stem Cells Int*. 2016;2016:1314709. doi:10.1155/2016/1314709
- 29. Wang Y, Yin P, Bian GL, et al. The combination of stem cells and tissue engineering: an advanced strategy for blood vessels regeneration and vascular disease treatment. *Stem Cell Research & Therapy*. 2017;8(1):194. doi:10.1186/s13287-017-0642-y
- 30. Barcena AJR, Dhal K, Patel P, Ravi P, Kundu S, Tappa K. Current Biomedical Applications of 3D-Printed Hydrogels. *Gels*. 2023;10(1). doi:10.3390/gels10010008
- 31. Priester C, MacDonald A, Dhar M, Bow A. Examining the Characteristics and Applications of Mesenchymal, Induced Pluripotent, and Embryonic Stem Cells for Tissue Engineering Approaches across the Germ Layers. *Pharmaceuticals (Basel)*. 2020;13(11). doi:10.3390/ph13110344

- 32. Schott NG, Friend NE, Stegemann JP. Coupling osteogenesis and vasculogenesis in engineered orthopedic tissues. *Tissue Engineering Part B: Reviews*. 2021;27(3):199-214. doi:10.1089/ten.teb.2020.0132. PMID:32854589
- 33. De Pieri A, Rochev Y, Zeugolis DI. Scaffold-free cell-based tissue engineering therapies: advances, shortfalls and forecast. *NPJ Regenerative Medicine*. 2021;6(1):18. doi:10.1038/s41536-021-00133-3. PMID:33782415
- 34. Brown PT, Handorf AM, Jeon WB, Li WJ. Stem cell-based tissue engineering approaches for musculoskeletal regeneration. *Curr Pharm Des*. 2013;19(19):3429-3445. doi:10.2174/13816128113199990350
- 35. Sanjurjo-Rodríguez C, Castro-Viñuelas R, Piñeiro-Ramil M, et al. Versatility of Induced Pluripotent Stem Cells (iPSCs) for Improving the Knowledge on Musculoskeletal Diseases. *Int J Mol Sci.* 2020;21(17). doi:10.3390/ijms21176124
- 36. Dennis SC, Berkland CJ, Bonewald LF, Detamore MS. Endochondral ossification for enhancing bone regeneration: converging native extracellular matrix biomaterials and developmental engineering in vivo. *Tissue Eng Part B Rev.* 2015;21(3):247-266. doi:10.1089/ten.TEB.2014.0419
- 37. Tripathi S, Mandal SS, Bauri S, Maiti P. 3D bioprinting and its innovative approach for biomedical applications. *MedComm* (2020). 2023;4(1):e194. doi:10.1002/mco2.194
- 38. Farooq M, Khan AW, Kim MS, Choi S. The Role of Fibroblast Growth Factor (FGF) Signaling in Tissue Repair and Regeneration. *Cells*. 2021;10(11). doi:10.3390/cells10113242
- 39. Li Y, Liu Y, Bai H, et al. Sustained Release of VEGF to Promote Angiogenesis and Osteointegration of Three-Dimensional Printed Biomimetic Titanium Alloy Implants. *Front Bioeng Biotechnol*. 2021;9:757767. doi:10.3389/fbioe.2021.757767
- 40. Sun XT, Ding YT, Yan XG, et al. Angiogenic synergistic effect of basic fibroblast growth factor and vascular endothelial growth factor in an in vitro quantitative microcarrier-based three-dimensional fibrin angiogenesis system. *World J Gastroenterol*. 2004;10(17):2524-2528. doi:10.3748/wjg.v10.i17.2524
- 41. Asahara T, Bauters C, Zheng LP, et al. Synergistic Effect of Vascular Endothelial Growth Factor and Basic Fibroblast Growth Factor on Angiogenesis In Vivo. *Circulation*. 1995;92(9):365-371. doi:10.1161/01.CIR.92.9.365

- 42. Shi M, Xia L, Chen Z, et al. Europium-doped mesoporous silica nanosphere as an immune-modulating osteogenesis/angiogenesis agent. *Biomaterials*. 2017;144:176-187. doi:10.1016/j.biomaterials.2017.08.027
- 43. Revete A, Aparicio A, Cisterna BA, et al. Advancements in the Use of Hydrogels for Regenerative Medicine: Properties and Biomedical Applications. *Int J Biomater*. 2022;2022:3606765. doi:10.1155/2022/3606765
- 44. Laiva AL, O'Brien FJ, Keogh MB. Innovations in gene and growth factor delivery systems for diabetic wound healing. *J Tissue Eng Regen Med*. 2018;12(1):e296-e312. doi:10.1002/term.2443
- 45. Beheshtizadeh N, Gharibshahian M, Bayati M, et al. Vascular endothelial growth factor (VEGF) delivery approaches in regenerative medicine. *Biomedicine & Pharmacotherapy*. 2023;166:115301. doi:10.1016/j.biopha.2023.115301
- 46. Flores-Rojas GG, Gómez-Lazaro B, López-Saucedo F, Vera-Graziano R, Bucio E, Mendizábal E. Electrospun Scaffolds for Tissue Engineering: A Review. *Macromol.* 2023;3(3):524-553. doi:10.3390/macromol3030031
- 47. Nazarnezhad S, Baino F, Kim HW, Webster TJ, Kargozar S. Electrospun Nanofibers for Improved Angiogenesis: Promises for Tissue Engineering Applications. *Nanomaterials (Basel)*. 2020;10(8). doi:10.3390/nano10081609
- 48. Oliveira ÉR, Nie L, Podstawczyk D, et al. Advances in growth factor delivery for bone tissue engineering. *International journal of molecular sciences*. 2021;22(2):903. doi:10.3390/ijms22020903. PMID:33477502
- 49. Krell ES, DiGiovanni CW. The Efficacy of Platelet-Derived Growth Factor as a Bone-Stimulating Agent. *Foot Ankle Clin*. 2016;21(4):763-770. doi:10.1016/j.fcl.2016.07.002
- 50. DiGiovanni CW, Lin SS, Daniels TR, et al. The Importance of Sufficient Graft Material in Achieving Foot or Ankle Fusion. *J Bone Joint Surg Am*. 2016;98(15):1260-1267. doi:10.2106/jbjs.15.00879
- 51. Kuttappan S, Mathew D, Jo JI, et al. Dual release of growth factor from nanocomposite fibrous scaffold promotes vascularisation and bone regeneration in rat critical sized calvarial defect. *Acta Biomater*. 2018;78:36-47. doi:10.1016/j.actbio.2018.07.050

- 52. Bouyer M, Guillot R, Lavaud J, et al. Surface delivery of tunable doses of BMP-2 from an adaptable polymeric scaffold induces volumetric bone regeneration. *Biomaterials*. 2016;104:168-181. doi:10.1016/j.biomaterials.2016.06.001
- 53. Naskar D, Ghosh AK, Mandal M, Das P, Nandi SK, Kundu SC. Dual growth factor loaded nonmulberry silk fibroin/carbon nanofiber composite 3D scaffolds for in vitro and in vivo bone regeneration. *Biomaterials*. 2017;136:67-85. doi:10.1016/j.biomaterials.2017.05.014
- 54. Udomluck N, Lee H, Hong S, Lee SH, Park H. Surface functionalization of dual growth factor on hydroxyapatite-coated nanofibers for bone tissue engineering. *Applied Surface Science*. 2020;520:146311. doi:10.1016/j.apsusc.2020.146311
- 55. Lee J, Lee S, Ahmad T, et al. Human adiposederived stem cell spheroids incorporating platelet-derived growth factor (PDGF) and bio-minerals for vascularized bone tissue engineering. *Biomaterials*. 2020;255:120192. doi:10.1016/j.biomaterials.2020.120192
- 56. Wang L, Fang M, Xia Y, et al. Preparation and biological properties of silk fibroin/nano-hydroxyapatite/graphene oxide scaffolds with an oriented channel-like structure. *RSC Advances*. 2020;10(17):10118-10128. doi:10.1039/C9RA09710D
- 57. Xu F, Dawson C, Lamb M, et al. Hydrogels for Tissue Engineering: Addressing Key Design Needs Toward Clinical Translation. *Front Bioeng Biotechnol*. 2022;10:849831. doi:10.3389/fbioe.2022.849831
- 58. Ducheyne P. *Comprehensive Biomaterials*. Vol 1. Elsevier; 2015. doi:<u>10.1016/</u> <u>B978-0-08-055294-1.00275-0</u>
- 59. Hu K, Olsen BR. The roles of vascular endothelial growth factor in bone repair and regeneration. *Bone*. 2016;91:30-38. doi:10.1016/j.bone.2016.06.013. PMID:27353702
- 60. Keceli HG, Bayram C, Celik E, Ercan N, Demirbilek M, Nohutcu RM. Dual delivery of platelet-derived growth factor and bone morphogenetic factor-6 on titanium surface to enhance the early period of implant osseointegration. *Journal of Periodontal Research*. 2020;55(5):694-704. doi:10.1111/jre.12756
- 61. Zhen R, Yang J, Wang Y, et al. Hepatocyte growth factor improves bone regeneration via the bone morphogenetic protein-2-mediated NF-KB signaling pathway. *Mol Med Rep.* 2018;17(4):6045-6053. doi:10.3892/mmr.2018.8559

- 62. Wallmichrath J, Stark GB, Kneser U, et al. Epidermal growth factor (EGF) transfection of human bone marrow stromal cells in bone tissue engineering. *J Cell Mol Med*. 2009;13(8b):2593-2601. doi:10.1111/j.1582-4934.2008.00600.x
- 63. Di Francesco D, Pigliafreddo A, Casarella S, Di Nunno L, Mantovani D, Boccafoschi F. Biological materials for tissue-engineered vascular grafts: Overview of recent advancements. *Biomolecules*. 2023;13(9):1389. doi:10.3390/biom13091389. PMID:37759789
- 64. Chen M, Jiang R, Deng N, Zhao X, Li X, Guo C. Natural polymer-based scaffolds for soft tissue repair. Mini Review. *Frontiers in Bioengineering and Biotechnology*. Published online July 19, 2022:10. doi:10.3389/fbioe.2022.954699
- 65. Liu Y, Nelson T, Chakroff J, et al. Comparison of polyglycolic acid, polycaprolactone, and collagen as scaffolds for the production of tissue engineered intestine. *J Biomed Mater Res B Appl Biomater*. 2019;107(3):750-760. doi:10.1002/jbm.b.34169
- 66. Bertsch C, Maréchal H, Gribova V, et al. Biomimetic bilayered scaffolds for tissue engineering: from current design strategies to medical applications. *Advanced healthcare materials*. 2023;12(17):2203115. doi:10.1002/adhm.202203115. PMID:36807830
- 67. Aishwarya B, Revathi V, Singh N, et al. Biodegradable Polymer Biomaterials for Tissue Engineering Applications: A Critical Review. Published online 2024:01051. doi:10.1051/e3sconf/202452901051
- 68. Zhu T, Zhou H, Chen X, Zhu Y. Recent advances of responsive scaffolds in bone tissue engineering. *Frontiers in Bioengineering and Biotechnology*. 2023;11:1296881. doi:10.3389/fbioe.2023.1296881. PMID:38047283
- 69. Loh QL, Choong C. Three-dimensional scaffolds for tissue engineering applications: role of porosity and pore size. *Tissue Eng Part B Rev*. 2013;19(6):485-502. doi:10.1089/ten.TEB.2012.0437
- 70. Preethi A, Bellare JR. Tailoring Scaffolds for Orthopedic Application With Anti-Microbial Properties: Current Scenario and Future Prospects. *Frontiers in Materials*. Published online February 18, 2021:7. doi:10.3389/fmats.2020.594686
- 71. Vallet-Regí M, Lozano D, González B, Izquierdo-Barba I. Biomaterials against bone infection. *Advanced healthcare materials*. 2020;9(13):2000310. doi:10.1002/adhm.202000310. PMID:32449317

- 72. Yuan X, Zhu W, Yang Z, et al. Recent Advances in 3D Printing of Smart Scaffolds for Bone Tissue Engineering and Regeneration. *Advanced Materials*. 2024;36(34):2403641. doi:10.1002/adma.202403641
- 73. Tatara AM, Mikos AG. Tissue Engineering in Orthopaedics. *J Bone Joint Surg Am*. 2016;98(13):1132-1139. doi:10.2106/jbjs.16.00299
- 74. Todd EA, Mirsky NA, Silva BLG, et al. Functional Scaffolds for Bone Tissue Regeneration: A Comprehensive Review of Materials, Methods, and Future Directions. *Journal of Functional Biomaterials*. 2024;15(10):280. doi:10.3390/jfb15100280. PMID:39452579
- 75. Ghassemi T, Shahroodi A, Ebrahimzadeh MH, Mousavian A, Movaffagh J, Moradi A. Current concepts in scaffolding for bone tissue engineering. *Archives of bone and joint surgery*. 2018;6(2):90.
- 76. Barakeh W, Zein O, Hemdanieh M, Sleem B, Nassereddine M. Enhancing Hip Arthroplasty Outcomes: The Multifaceted Advantages, Limitations, and Future Directions of 3D Printing Technology. *Cureus*. 2024;16(5):e60201. doi:10.7759/cureus.60201
- 77. Tremblay PL, Hudon V, Berthod F, Germain L, Auger FA. Inosculation of tissue-engineered capillaries with the host's vasculature in a reconstructed skin transplanted on mice. *American journal of transplantation*. 2005;5(5):1002-1010. doi:10.1111/j.1600-6143.2005.00790.x
- 78. Mercado-Pagán ÁE, Stahl AM, Shanjani Y, Yang Y. Vascularization in bone tissue engineering constructs. *Annals of biomedical engineering*. 2015;43:718-729. doi:10.1007/s10439-015-1253-3. PMID:25616591
- 79. Folkman J, Hochberg M. Self-regulation of growth in three dimensions. *The Journal of experimental medicine*. 1973;138(4):745-753. doi:10.1084/jem.138.4.745
- 80. Devillard CD, Marquette CA. Vascular tissue engineering: challenges and requirements for an ideal large scale blood vessel. *Frontiers in Bioengineering and Biotechnology*. 2021;9:721843. doi:10.3389/fbioe.2021.721843. PMID:34671597
- 81. Hegen A, Blois A, Tiron CE, et al. Efficient in vivo vascularization of tissue-engineering scaffolds. *Journal of tissue engineering and regenerative medicine*. 2011;5(4):e52-e62. doi:10.1002/term.336. PMID:20865694

- 82. Song HHG, Rumma RT, Ozaki CK, Edelman ER, Chen CS. Vascular tissue engineering: progress, challenges, and clinical promise. *Cell stem cell*. 2018;22(3):340-354. doi:10.1016/j.stem.2018.02.009. PMID:29499152
- 83. Deb S, Mandegaran R, Di Silvio L. A porous scaffold for bone tissue engineering/45S5 Bioglass® derived porous scaffolds for co-culturing osteoblasts and endothelial cells. *Journal of Materials Science: Materials in Medicine.* 2010;21:893-905.
- 84. Drury JL, Mooney DJ. Hydrogels for tissue engineering: scaffold design variables and applications. *Biomaterials*. 2003;24(24):4337-4351. doi:10.1016/S0142-9612(03)00340-5
- 85. Chen S, Zhou X, Li T, He C. Vascularization and Innervation for Bone Tissue Engineering. *Accounts of Materials Research*. 2024;5(9):1121-1133. doi:10.1021/accountsmr.4c00165

- 86. Kazemnejad S, Khanmohammadi M, Baheiraei N, Arasteh S. Current State of Cartilage Tissue Engineering using Nanofibrous Scaffolds and Stem Cells. *Avicenna J Med Biotechnol*. 2017;9(2):50-65.
- 87. Apelgren P, Amoroso M, Säljö K, et al. Vascularization of tissue engineered cartilage Sequential in vivo MRI display functional blood circulation. *Biomaterials*. 2021;276:121002. doi:10.1016/j.biomaterials.2021.121002
- 88. Higgins JP, Borumandi F, Bürger HK, et al. Nonvascularized Cartilage Grafts Versus Vascularized Cartilage Flaps: Comparison of Cartilage Quality;Months After Transfer. *Journal of Hand Surgery*. 2018;43(2):188.e1-188.e8. doi:10.1016/j.jhsa.2017.09.014