

General

Hip Osteoarthritis and the Human Microbiome: Current Concepts, Biological Insights and Future Directions

Augusto Ferrini^{1,2} , Alessandro Del Monaco^{1,2}, Biagio Zampogna^{2,3,4}, Antonio Bosso^{1,2}, Andrea Zampoli^{1,2}, Francesco R. Parisi^{1,2}, Chiara Capperucci^{1,2}, Lorenzo A. Diaz Balzani^{1,2}, Rocco Papalia^{1,2}

¹ Research Unit of Orthopaedic and Trauma Surgery, Department of Medicine and Surgery, Università Campus Bio-Medico, ² Operative Research Unit of Orthopaedic and Trauma Surgery, Policlinico Universitario Campus Bio Medico, ³ Department of Medicine and Surgery, Università Campus Bio-Medico, ⁴ BIOMORF Department of Biomedical, Dental, Morphological and Functional Images, University of Messina

Keywords: Hip osteoarthritis, Gut microbiome, Dysbiosis, Gut-joint axis, Inflammation, Microbiota-targeted therapy

<https://doi.org/10.52965/001c.164467>

Orthopedic Reviews

Vol. 18, 2026

Osteoarthritis (OA) is among the leading causes of disability worldwide, yet its pathogenesis remains incompletely understood. Once considered a “non-inflammatory” degenerative disorder, OA is now recognized as a condition driven by chronic low-grade inflammation. Emerging evidence implicates gut dysbiosis as a modifiable risk factor, promoting systemic inflammation through impaired gut permeability and translocation of microbial components. These immune-modulating molecules can trigger pro-inflammatory cascades and pathological bone remodeling. This review summarizes current knowledge linking gut dysbiosis and knee osteoarthritis and extends these insights to hip osteoarthritis (HOA). Observational and genetic studies support a causal role for the microbiota, identifying specific taxa associated with either increased or reduced HOA risk. Preclinical and clinical data describe a mechanistic axis linking intestinal dysbiosis, synovial inflammation, and cartilage degeneration. In animal models, particularly under high-fat/high-sucrose diets, visceral adiposity emerges as a major driver of joint damage. While microbial metabolites such as short-chain fatty acids appear protective, the detection of microbial DNA within joint tissues remains controversial, suggesting possible joint-specific microbial ecosystems. Based on these findings, microbiota-targeted strategies are under investigation as potential interventions to influence OA progression and relieve hip pain. However, longitudinal cohorts and randomized clinical trials in HOA are needed to clarify causal mechanisms and therapeutic efficacy in HOA.

1. INTRODUCTION

Osteoarthritis (OA) represents one of the leading causes of disability and functional impairment among working-age adults and the elderly population.¹ According to the Global Burden of Disease Study 2019, the global prevalence of OA has dramatically increased by 113.25%, rising from 247.51 million in 1990 to 527.81 million in 2019.² OA represents a huge challenge for healthcare systems, accounting for an estimated 1-2.5% of the gross national product in developed countries, and projected to quadruple by 2030.³ Within OA phenotypes, Hip Osteoarthritis (HOA) is the second most common subtype after knee OA (KOA), and is characterized by progressive cartilage loss, subchondral bone remodeling, and synovial inflammation.⁵ Although risk factors including biomechanical stress, advanced age, obesity, and altered joint loading are well-established, many individuals exposed to these factors do not develop OA.^{6,7} In the absence of secondary causes such as femoroacetabular impingement (FAI), post-traumatic arthritis, rheumatoid arthritis, or osteonecrosis of the

femoral head, many HOA cases remain idiopathic. This strongly suggests that systemic mechanisms contribute to disease initiation and progression.⁸ Traditionally considered a “non-inflammatory” degenerative disorder, OA is now increasingly recognized as a condition characterized by chronic low-grade inflammation.⁹ Recent evidence suggested that this systemic inflammatory state may be influenced by the gut microbiome, defined as the collective genome and metabolic products of trillions of intestinal microorganisms.^{10,11} Dysbiosis, or disruption of the gut microbial balance, may compromise intestinal permeability, leading to systemic translocation of bacteria components such as lipopolysaccharides (LPS) and short-chain fatty acids (SCFAs). These molecules may stimulate immune responses, trigger pro-inflammatory cascades, and ultimately promote pathological bone remodeling.¹²⁻¹⁴ The role of gut-joint axis has been progressively acknowledged in OA, where altered microbial composition has been associated with increased synovial inflammation, pain sensitization, and accelerated joint degeneration.¹⁵ Given the pathophysiological features among OA phenotypes, similar microbiome-related mechanisms may contribute to idio-

pathic HOA, potentially influencing immune responses, cartilage homeostasis, and subchondral bone turnover, despite its deeper anatomical location. Increasing our knowledge of the gut microbiota and the role it may play in the pathogenesis of HOA could improve our understanding of disease mechanism, enable a more personalized risk assessment, and open the way to new therapeutic strategies and approaches for management and prevention of OA. While the “gut–joint axis” has been primarily characterized in knee osteoarthritis, this review extends the concept to the hip joint, introducing the notion of a “gut–hip axis” to describe the potential site-specific manifestation of systemic microbial influences. Due to the exploratory nature of the subject and the lack of hip-specific evidence, this manuscript was conducted as a narrative review and does not include a systematic search, PRISMA flow diagram, or formal risk-of-bias evaluation. To our knowledge, this is among the first review specifically focusing on the emerging link between dysbiosis and HOA, aiming to explore current evidence and outline future research directions in this promising and evolving field.

2. METHODS

This narrative review was conducted to summarize the current evidence regarding relationship between the gut microbiota dysbiosis and HOA. A literature search was performed using PubMed, Scopus and Web of Science databases. The following search terms and combinations were used: “hip osteoarthritis”, “osteoarthritis”, “gut microbiome”, “gut microbiota”, “dysbiosis”, “gut–joint axis”, “gut–hip axis”, “short-chain fatty acids”, “inflammation”, and “microbiota-targeted therapy”. Relevant preclinical and clinical studies, systematic reviews, and randomized controlled trials were considered. Given the limited HOA-specific evidence available, studies focusing on KOA models and broader gut–joint axis were also included to support the biological rationale of this review. Only articles published in English and considered relevant to the scope of the review were included.

3. PATHOPHYSIOLOGY OF HIP OSTEOARTHRITIS

The pathogenesis of early HOA is complex, multifactorial, and not yet fully elucidated. Physiological loading is essential for maintaining joint tissue homeostasis,^{16,17} but excessive or abnormal mechanical stress disrupts the balance between synthesis and degradation, leading to progressive cartilage degeneration.^{18–21} Repetitive shear stress induces a reduction in type II collagen and proteoglycan synthesis, an increase in pro-inflammatory mediators, and chondrocyte apoptosis.²¹ Within the osteochondral unit, subchondral bone remodeling with increased porosity and microcracks enables biochemical and mechanical communication between cartilage and bone.^{22–24} In this bidirectional exchange, cartilage releases cytokines and osteoclastogenic factors,^{22,25} while subchondral osteoblasts contribute to

cartilage degradation.²⁶ Early synovitis²⁷ further supports the modern concept of OA as a whole-joint disease from its earliest stages. From an etiological perspective, HOA arises from both local, joint-specific factors and systemic influences that modulate the biological response to mechanical stress. Local morphological abnormalities alter joint load distribution and predispose to tissue degeneration. In developmental dysplasia of the hip (DDH), a shallow acetabulum increases load on the acetabular rim, and cartilage injury.²⁰ In FAI, cam deformities induce anterosuperior delamination, while pincer variants cause labral damage due to acetabular over coverage.^{28,29} These morphological changes exist along a continuum, with severe deformities linked to early-onset OA and milder variants associated with primary late-onset OA.³⁰ Periarticular muscle weakness and labral tears further exacerbate abnormal joint loading, accelerating cartilage degeneration.^{31–34} Systemic factors also influence joint susceptibility. Aging reduces tissue repair capacity,^{35–38} while hormonal changes may explain sex-specific trend: HOA is more frequent in men under 50 years but rises in postmenopausal women, with estrogen therapy may exert protective effects.^{18,39} While occupational exposure to heavy manual labor, particularly farming, increases the risk of HOA in predisposed individuals, no consistent association has been found with sports participation.^{40,41} Twin studies have shown that genetic factors may account for up to 60% of HOA risk,⁴² and ethnic differences suggest that morphological and genetic variations may be protective in some populations.⁴³ Obesity represents a dual contributor: each 5-unit increase in BMI is associated with an approximately 11% increase in HOA incidence.⁴⁴ Beyond mechanical overload, adiposity contributes to chronic low-grade inflammation, through adipokines and cytokines, affecting cartilage and bone metabolism.⁴⁵ The link between obesity and OA in non-weight-bearing joint, such as the hands, further supports a systemic inflammation mechanism.⁴⁶

4. THE GUT–JOINT AXIS AND KOA: A PARADIGM FOR UNDERSTANDING HOA

Idiopathic OA is no longer regarded as a purely mechanical disorder but as a disease of chronic low-grade inflammation. Even before overt structural damage, chondrocytes exposed to chronic inflammatory and metabolic stress shift toward glycolysis, increasing reactive oxygen species, suppressing autophagy, and upregulating matrix-degrading enzymes.⁴⁷ Synovial macrophages in OA show an imbalance toward the pro-inflammatory M1 phenotype, driving tissue degradation and joint pain. Cytokines such as IL-6 and chemokines like CCL2, along with transcriptional regulators including NF- κ B and LEF1, coordinate the catabolic and neurogenic responses that characterize the OA microenvironment.⁴⁸ RNA-binding proteins (RBPs) have emerged as post-transcriptional regulators of inflammation, matrix remodeling, and chondrocyte senescence.⁴⁹ Extracellular vesicles (EVs) further amplify degeneration by transferring cytokines, MMPs, and microRNAs between chondrocytes, synoviocytes, and joint-resident cells pro-

moting apoptosis, and synovial activation.⁵⁰ Over the last decade, a growing body of literature has emphasized the importance of the gut-joint axis, highlighting the potential impact of the intestinal microbiome on musculoskeletal tissues. Evidence from both preclinical and clinical studies has outlined a cascade linking intestinal dysbiosis, impaired barrier permeability, systemic dissemination of microbial products, immune activation, synovial inflammation, and pain sensitization. Most of the mechanistic and clinical evidence currently available derives from knee osteoarthritis (KOA) cohorts or animal KOA models. Therefore, the following concepts should be considered as extrapolations rather than confirmed hip-specific findings. Although both HOA and KOA share similar inflammatory pathways, anatomical and structural differences may influence the impact of microbial-derived inflammation. The hip joint demonstrates increased subchondral bone thickness and elevated microstructural porosity relative to the knee, which may promote microbial product transport and amplify subchondral immune activity.^{22,23} Moreover, the deeper vascular supply of the femoral head may increase exposure to circulating microbial metabolites or endotoxins in conditions of systemic dysbiosis. On the other hand, the knee joint may exhibit a quicker and more dynamic inflammatory response due to its higher synovial volume and more superficial vascular.^{25,51} These differences may explain subtle variations in how microbial-driven inflammation contributes to cartilage degeneration in HOA versus KOA.

4.1. EVIDENCE FROM ANIMAL MODELS OF DYSBIOSIS

Animal models have provided some of the clearest mechanistic evidence linking dysbiosis to joint degeneration. Exposure to high fat/high sucrose (HFS) diets induces profound alterations in gut microbiota, characterized by loss of commensal *Lactobacillus* spp. and enrichment of pro-inflammatory *Methanobrevibacter* spp. Sun et al expanded these observations by showing that high-fat diet (HFD) compromises epithelial integrity, reduces tight junction proteins expression, such as zonula occludens 1 (ZO-1), and facilitates the translocation of LPS. In addition, bacterial outer-membrane vesicles (OMVs) may bypass mucosal defenses, delivering pro-inflammatory signals into the sub-mucosa.⁵² Notably, visceral adiposity rather than overall body weight represents the strongest predictor of cartilage damage, highlighting the contribution of adipose-derived inflammatory mediators, including leptin, GRO-KC, MIP-1, IP-10, MIP-2, in both serum and synovial fluid. Increased intra-articular IL-1 α levels further correlate with structural cartilage damage.^{53,54} These findings support the concept that dysbiosis is not merely an association but an active driver of joint pathology. Although animal studies are useful to explore mechanisms, most models induce dysbiosis with extreme diets, use very small sample sizes, and do not replicate typical human HOA exposures. These factors may limit internal validity and raise questions about external translation to human hip OA.

4.2. MICROBIAL METABOLITES AND BARRIER INTEGRITY

A central mechanism involves microbial metabolites, particularly short chain fatty acids (SCFAs), produced through microbial fermentation of nondigestible dietary fibers. Propionate and butyrate, generated by specialized taxa including *Akkermansia muciniphila*, *Eubacterium dolichum*, *Ruminococcus bromii*, *Bacteroides* spp, exerted protective effects on bone and joint tissues.⁵⁵⁻⁵⁸ SCFAs inhibit osteoclast differentiation by reducing pro-inflammatory cytokines including TNF- α , IL-1 β , and IL-6, as well as down-regulating RANKL expression by B cells.^{59,60} Additionally, butyrate enhances osteoblast differentiation via T regulatory cells and stimulation of Wnt10b signaling.^{61,62} Furthermore, SCFAs contribute to maintain epithelial barrier integrity by reducing systemic inflammation and microbial translocation, thereby indirectly protecting against bone loss in inflammatory and postmenopausal osteoporosis.^{60,63,64} Ovariectomized mice analyses confirmed the potential of microbiota-targeted therapies by showing that *Lactobacillus* strains supplementation increases Claudin family protein expression, improves intestinal permeability, promotes osteoblast activity, and prevents sex steroid-induced bone loss.^{65,66} Preclinical and clinical researches have supported the important role that the microbiome plays in regulating skeletal homeostasis and contributing to bone-related diseases.^{60,67} He et al.⁶⁸ demonstrated that decreased SCFA-producing taxa in postmenopausal women are associated with lower bone mineral density and higher bone resorption. In parallel, butyrate might modulate zonulin, a physiologic regulator of gut permeability. Increased permeability occurred in celiac disease or after bacterial stimuli. Guido et al. showed that reduction in the *Bacteroidetes* phylum impairs gut permeability. In this context, an abundance of *Streptococcus* spp increased LPS exposure, activates macrophages via NF- κ B and the MAPK pathways, increases the M1/M2 macrophages ratio, and promotes production of MMPs and pro-inflammatory mediators. Chemokines such as the CCL2/CCR2 axis seem to be involved in the development of pain, while inflammation-induced upregulation of aquaporins (AQP1, AQP3) may trigger joint swelling in early OA, proteoglycans loss, and chondrocyte apoptosis.⁶⁹ Such evidence supports the role of the gut microbiome in preserving bone health and modulating disease-related responses.

4.3. IMMUNE MECHANISMS AND INFLAMMATORY RESPONSE

An impaired gut barrier allows the translocation of microbe-associated molecular patterns (MAMPs), including LPS, peptidoglycan (PGN), flagellin, and bacterial DNA, which are able to activate innate immunity in distant tissues. By binding to pattern recognition receptors, especially Toll-like receptors (TLRs) and nucleotide-binding oligomerization domain (NOD)-like receptors (NLRs), these molecules stimulate pro-inflammatory cascades in cartilage, bone, and synovium. Moreover, PGN may trigger NLRP3 inflammasome activation, leading to macrophages

release of IL-1 β , and impaired cartilage extracellular matrix synthesis. Similarly, LPS engage the LPS-LBP-CD14-TLR4-MD-2 complexes, promotes NF- κ B activation and upregulates TNF, IL-1 β , IL-6, IL-8, RANKL, and MMPs, ultimately accelerating cartilage degradation.

In addition, LPS may induce inflammation in adipose tissue, eventually stimulating the chronic inflammation state inside the knee joint.⁷⁰⁻⁷⁵ Gut permeability may also lead to transient bacteremia, whereby microbes or microbial products reach the joint either directly or via immune cells acting as “Trojan Horse”, as suggested by a recent systematic review.⁵³

4.4. GUT MICROBIOME AND OSTEOARTHRITIS

The presence of microbial DNA within joint tissues remains among the most debated findings. Dunn et al. identified distinct microbial communities in cartilage, with KOA samples enriched in Firmicutes and HOA characterized by the Proteobacteria.⁷⁶ Similarly, other studies reported increased Clostridiales and decreased Lactobacillus spp. in OA joints.⁵³ Although contamination cannot be excluded, the consistency of findings suggests the hypothesis of a resident joint microbiome. This concept remains highly debated. Because joint and cartilage samples are extremely low-biomass tissues, the detection of microbial DNA is inherently vulnerable to contamination from reagents, laboratory workflow, and environmental sources. Interestingly, Bonanzinga et al. reported that Cutibacterium, Staphylococcus, and Paracoccus spp. seem to be dominant in non-OA samples, whereas OA joints show higher Proteobacteria abundance.⁷⁷ The divergence between hip and knee findings is particularly relevant, suggesting joint-specific microbial ecosystems that may differentially modulate disease pathogenesis.

The gut–joint axis is increasingly recognized as a potential driver of the pathogenesis of OA. As detailed above, SCFAs and other microbial metabolites may mediate protective effects by modulating inflammation and barrier integrity.⁷⁸ Some microbial antigens may reach deep cartilage layers, triggering innate immune responses.^{79,80} Other microbial products may also influence chondrocyte epigenetics, altering DNA methylation and non-coding RNA profiles, especially in obesity-related OA.⁸¹ The microbiota influences systemic aging, oxidative stress, and circadian rhythms, all of which are implicated in joint degeneration⁸²; moreover, circadian misalignment alters microbial composition, while dysbiosis reciprocally impairs circadian regulation, thereby disrupting cartilage homeostasis.^{83,84} Beyond structural damage, dysbiosis also appears to modulate nociception. Data from the Rotterdam Study showed that the abundance of Streptococcus spp. correlates with increased WOMAC pain scores and MRI-detected effusion in patients with KOA.¹⁰ Recent systematic review supported the clinical association between pain severity and microbiome alterations in KOA.⁷⁷ Furthermore, systemic inflammatory mediators, such as IL-6, MDC, and IP-10, are significantly associated with pain in both KOA and HOA.⁵¹

Although these findings have yet to be replicated in HOA, they raise the possibility of shared microbial influ-

ences across joint sites.⁷⁶ Dietary findings reinforce these mechanisms.⁵⁴ Similarly, Schott et al. found that in murine model of OA induced by medial meniscal destabilization, a high-fat diet leads to gut dysbiosis, increased intestinal permeability, and elevated systemic IL-1 β and TNF- α levels, which accelerate cartilage degeneration.⁸⁵ Treatment with the prebiotic oligofructose supplementation restores a healthier microbiota, reduces circulating LPS, and attenuates joint damage, without significant weight loss, suggesting microbiota modulation as a potential chondroprotective strategy independent of mechanical stress.

4.5. GUT MICROBIOME AND HIP OSTEOARTHRITIS

Evidence from genetics, animal models, and observational studies supported a causal role of the gut microbiota in the pathogenesis of HOA. Mendelian randomization analyses identified specific genera such as Gordonibacter and Eubacterium (brachy group) as risk enhancers, while Senegalimassilia, Slackia, and Streptococcus are linked to reduced risk.⁸⁶ Animal studies provided further insights. In a canine model of spontaneous OA, Cintio et al. observed decreased fecal alpha diversity, increased Megamonas and decreased Paraprevotellaceae, Porphyromonadaceae, and Mogibacterium, findings consistent with human studies.⁸⁷ These changes may either drive or result from the OA process. Genetic approaches further underscored the causal links between microbiota and HOA. Li et al., using a two-step bidirectional Mendelian randomization strategy, identified a protective effect of Actinobacteria (notably Bifidobacteria) on OA risk, partially mediated through basal metabolic rate (BMR).⁸⁸ Elevated BMR was associated with increased OA risk, providing a novel mechanistic link between microbiota composition, systemic metabolism, and joint health. Conversely, Lee et al found no significant associations between gut microbiota and OA after multiple testing corrections, citing methodological limitations such as reliance on fecal data and joint-site heterogeneity.⁸⁹ Although Mendelian randomization analyses have been interpreted as supportive of a causal relationship between gut dysbiosis and OA, these findings must be considered with caution. In microbiome Mendelian studies in particular, weak-instrument bias, and multiple testing across numerous taxa may lead to overestimation of causality. Therefore, Mendelian randomization supports biological plausibility rather than definitive causality for hip OA. Beyond systemic effects, the concept of a joint-specific microbiome has gained traction. Goswami et al. showed that microbial compositions derived from OA joints differ from environmental and skin flora, even after intra-articular injections or surgery.⁹⁰ The bacterial profiles identified in their study were distinct from those commonly found in the skin, not attributable to procedural contamination, supporting the concept of a resident intra-articular microbiome.⁹⁰ Similarly, Fernandez-Rodriguez et al. demonstrated joint-specific microbial profiles in human knees, suggesting potential extension to the hip.⁹¹ These host-microbiota interactions may influence both the pathogenesis of OA and susceptibility to periprosthetic joint infections (PJI). Dunn et al. also identified distinct microbial DNA in hip cartilage, including

Pseudomonas, *Acinetobacter*, and *Streptococcus*, with joint-specific differences in microbial profiles and immune marker correlations.⁷⁶ These findings suggest that the microbiota may modulate local inflammation and cartilage integrity directly within the hip joint. Microbiome-host interactions have also been observed in obesity-related HOA. Anderson et al. found that obese patients undergoing anterior hip approach surgery have higher prevalence of *Enterobacteriales* in the skin microbiota overlying the surgical site compared to normal-weight individuals (21.4% vs. 2.8%).⁹² Such alterations may contribute to increased PJI risk in obese individuals, highlighting another microbiota-related pathway of surgical vulnerability. The therapeutic potential of the gut microbiota is emerging as a promising strategy in the management of OA. In a double-blind, randomized controlled trial, Karim et al. demonstrated that 12 weeks of high dose multistrain probiotics supplementation significantly improved pain, WOMAC scores, and postural balance, correlating with reduced plasma zonulin.⁹³ Similarly, in a larger trial involving 537 patients, Lei et al. demonstrated that 6 months of *Lactobacillus casei* Shirota supplementation improved WOMAC and VAS scores and reduced systemic inflammation (CRP levels).⁹⁴ The growing interest in this field has prompted authors such as Pedersini et al. to outline a trial protocol for advanced HOA and KOA (Kellgren–Lawrence ≥ 3), although clinical outcome data are not yet available.⁹⁵ Human evidence directly linking dysbiosis to hip osteoarthritis is still limited. Only a few studies have specifically investigated HOA, and their findings should be interpreted as preliminary. The available data suggest possible correlations, but the strength of evidence is currently insufficient to establish causality. These ongoing investigations will clarify whether microbiota-based therapies can be translated in clinical opportunity. [Figure 1.](#)

5. THERAPEUTIC IMPLICATIONS

Given these mechanistic links, microbiota-targeted strategies have been explored as potential therapeutic interventions. Synbiotics combinations, including oligofructose-inulin, *Lactobacillus rhamnosus*, and *Bifidobacterium lactis*, have been shown to restore beneficial taxa, increase tight junction protein expression, and reduce pro-inflammatory cytokines.⁹⁶ Prebiotics such as oligofructose may also exert prophylactic effects on OA development, by reducing gut permeability, increasing the expression of tight junction proteins, and modulating macrophage infiltration into synovium.⁵² Collectively, these approaches highlight the translational potential of manipulating the gut microbiota to modulate OA pathogenesis. Given the shared risk factors between KOA and HOA, including aging, obesity, metabolic syndrome, and low-grade systemic inflammation, it is plausible that similar microbiome-mediated pathways contribute to hip disease. To date, these mechanistic pathways have not been systematically linked to hip-specific outcomes. Thus, KOA serves as a blueprint for extending these concepts.

6. FUTURE DIRECTIONS

The gut-bone axis is emerging as a promising frontier in understanding the pathogenesis of HOA. However, most evidence comes from KOA or non-joint-specific microbiome studies, and true HOA-focused datasets are sparse. Therefore, mechanisms discussed in this review should be interpreted as biologically plausible but not fully demonstrated. Specific microbial taxa, metabolites, and functional pathways involved in cartilage degeneration and subchondral bone remodeling may be identified through high-resolution microbiome profiling and immunologic analyses. Randomized controlled trials are urgently required to evaluate microbiota-targeted interventions, such as well-characterized probiotics, dietary regimens promoting SCFA production, synbiotic, and, in selected cases, fecal microbiota transplantation.

7. CONCLUSION

Hip OA remains a debilitating condition with complex and incompletely understood pathophysiology. Traditional risk factors such as age, mechanical load, and structural abnormalities only partially explain its onset and progression. Although most of the recent advances in microbiome research have been achieved in KOA, the strong pathophysiological overlap between these two major forms of the disease suggests that dysbiosis may also play a central role in hip joint. Emerging findings support the concept that dysbiosis could represent a systemic, modifiable risk factor for OA, opening new avenues for prevention and treatment. Microbiome profiling could help identify specific subgroups of patients, particularly those without conventional risk factors, who may benefit from targeted interventions. In this context, microbiota-modulating strategies already explored in KOA, including probiotics, and SCFAs-enhancing diets, may eventually be evaluated in HOA as a potential adjunctive approach to slow disease progression. Overall, although current evidence remains preliminary, the close analogy with KOA strongly supports the existence of a gut-hip axis. The concept of a gut–hip axis remains promising but unproven. Confirming and expanding these findings in HOA may significantly advance our understanding of its pathogenesis and open the way for innovative therapeutic perspectives.

AUTHOR CONTRIBUTIONS

Conceptualization, A.F., A.D.M., L.A.D.B.; methodology, A.F., A.D.M., and L.A.D.B., A.B.; software, A.B. and A.Z.; validation, B.Z., and L.A.D.B.; investigation, A.F., A.D.M. and A.B.; resources, F.R.P., A.Z., and C.C.; writing—original draft preparation, A.F., A.D.M., A.B.; writing—review and editing, F.R.P., A.Z., and C.C.; visualization, B.Z.; supervision, L.A.D.B., E.A., R.P.; All authors have read and agreed to the published version of the manuscript

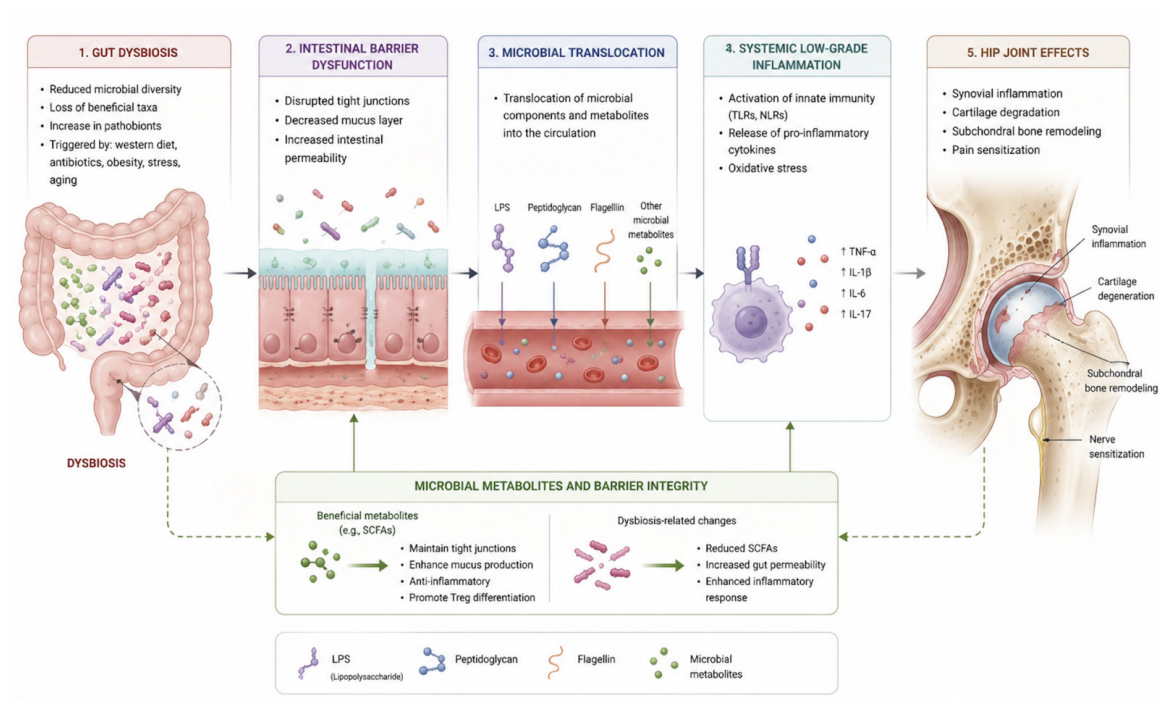


Figure 1. Schematic representation of the gut–joint axis in hip osteoarthritis. Gut dysbiosis may promote systemic inflammation and contribute to the development and progression of hip osteoarthritis.

FUNDING

This research received no external funding.

INSTITUTIONAL REVIEW BOARD STATEMENT

Not applicable.

INFORMED CONSENT STATEMENT

Not applicable.

DATA AVAILABILITY STATEMENT

Not applicable.

CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

Submitted: May 30, 2026 EDT. Accepted: June 25, 2026 EDT.

Published: July 10, 2026 EDT.

REFERENCES

1. Lawrence RC, Felson DT, Helmick CG, et al. Estimates of the Prevalence of Arthritis and Other Rheumatic Conditions in the United States, Part II. *Arthritis Rheum.* 2008;58:26-35. doi:[10.1002/art.23176](https://doi.org/10.1002/art.23176)
2. He Y, Jiang W, Wang W. Global Burden of Osteoarthritis in Adults Aged 30 to 44 Years, 1990 to 2019: Results from the Global Burden of Disease Study 2019. *BMC Musculoskelet Disord.* 2024;25:303. doi:[10.1186/s12891-024-07442-w](https://doi.org/10.1186/s12891-024-07442-w)
3. Leifer VP, Katz JN, Losina E. The Burden of OA-Health Services and Economics. *Osteoarthritis and Cartilage.* 2022;30:10-16. doi:[10.1016/j.joca.2021.05.007](https://doi.org/10.1016/j.joca.2021.05.007)
4. Ferrini A, He M, Anwar K, Papalia R, Tuncay I, Parvizi J. Total Knee Arthroplasty: Have We Made Any Strides over the Last 50 Years? *Knee Surg Sports Traumatol Arthrosc.* Published online 2025. doi:[10.1002/ksa.70073](https://doi.org/10.1002/ksa.70073)
5. Fan Z, Yan L, Liu H, et al. The Prevalence of Hip Osteoarthritis: A Systematic Review and Meta-Analysis. *Arthritis Res Ther.* 2023;25:51. doi:[10.1186/s13075-023-03033-7](https://doi.org/10.1186/s13075-023-03033-7)
6. Aresti N, Kassam J, Nicholas N, Achan P. Hip Osteoarthritis. *BMJ.* 2016;i3405. doi:[10.1136/bmj.i3405](https://doi.org/10.1136/bmj.i3405)
7. Hao X, Shang X, Liu J, Chi R, Zhang J, Xu T. The Gut Microbiota in Osteoarthritis: Where Do We Stand and What Can We Do? *Arthritis Res Ther.* 2021;23:42. doi:[10.1186/s13075-021-02427-9](https://doi.org/10.1186/s13075-021-02427-9)
8. Hunter DJ, Bierma-Zeinstra S. Osteoarthritis. *The Lancet.* 2019;393:1745-1759. doi:[10.1016/S0140-6736\(19\)30417-9](https://doi.org/10.1016/S0140-6736(19)30417-9)
9. van den Bosch MHJ, Blom AB, van der Kraan PM. Inflammation in Osteoarthritis: Our View on Its Presence and Involvement in Disease Development over the Years. *Osteoarthritis and Cartilage.* 2024;32:355-364. doi:[10.1016/j.joca.2023.12.005](https://doi.org/10.1016/j.joca.2023.12.005)
10. Boer CG, Radjabzadeh D, Medina-Gomez C, et al. Intestinal Microbiome Composition and Its Relation to Joint Pain and Inflammation. *Nat Commun.* 2019;10:4881. doi:[10.1038/s41467-019-12873-4](https://doi.org/10.1038/s41467-019-12873-4)
11. Almeida A, Mitchell AL, Boland M, et al. A New Genomic Blueprint of the Human Gut Microbiota. *Nature.* 2019;568:499-504. doi:[10.1038/s41586-019-0965-1](https://doi.org/10.1038/s41586-019-0965-1)
12. Heckmann ND, Culler MW, Mont MA, Lieberman JR, Parvizi J. Emerging Concepts in Periprosthetic Joint Infection Research: The Human Microbiome. *The Journal of Arthroplasty.* 2025;40:1821-1826. doi:[10.1016/j.arth.2025.01.001](https://doi.org/10.1016/j.arth.2025.01.001)
13. Yan J, Herzog JW, Tsang K, et al. Gut Microbiota Induce IGF-1 and Promote Bone Formation and Growth. *Proc Natl Acad Sci U S A.* 2016;113:E7554-E7563. doi:[10.1073/pnas.1607235113](https://doi.org/10.1073/pnas.1607235113)
14. Ferrini A, He M, Mont MA, Goodman SB, Parvizi J. Osteonecrosis of the Femoral Head: A Dysbiotic Condition? *J Arthroplasty.* Published online 2025. doi:[10.1016/j.arth.2025.11.044](https://doi.org/10.1016/j.arth.2025.11.044)
15. Huang Z, Chen J, Li B, et al. Faecal Microbiota Transplantation from Metabolically Compromised Human Donors Accelerates Osteoarthritis in Mice. *Annals of the Rheumatic Diseases.* 2020;79:646-656. doi:[10.1136/annrheumdis-2019-216471](https://doi.org/10.1136/annrheumdis-2019-216471)
16. Guilak F, Fermor B, Keefe FJ, et al. The Role of Biomechanics and Inflammation in Cartilage Injury and Repair. *Clin Orthop Relat Res.* Published online 2004;17-26. doi:[10.1097/01.blo.0000131233.83640.91](https://doi.org/10.1097/01.blo.0000131233.83640.91)
17. Setton LA, Mow VC, Müller FJ, Pita JC, Howell DS. Mechanical Behavior and Biochemical Composition of Canine Knee Cartilage Following Periods of Joint Disuse and Disuse with Remobilization. *Osteoarthritis Cartilage.* 1997;5:1-16. doi:[10.1016/S1063-4584\(97\)80027-1](https://doi.org/10.1016/S1063-4584(97)80027-1)
18. Eyre DR. Collagens and Cartilage Matrix Homeostasis. *Clin Orthop Relat Res.* Published online 2004;S118-122. doi:[10.1097/01.blo.0000144855.48640.b9](https://doi.org/10.1097/01.blo.0000144855.48640.b9)
19. Guilak F. Biomechanical Factors in Osteoarthritis. *Best Pract Res Clin Rheumatol.* 2011;25:815-823. doi:[10.1016/j.berh.2011.11.013](https://doi.org/10.1016/j.berh.2011.11.013)
20. Lane NE, Lin P, Christiansen L, et al. Association of Mild Acetabular Dysplasia with an Increased Risk of Incident Hip Osteoarthritis in Elderly White Women: The Study of Osteoporotic Fractures. *Arthritis Rheum.* 2000;43(2):400-404. doi:[10.1002/1529-0131\(200002\)43:2](https://doi.org/10.1002/1529-0131(200002)43:2)
21. Smith RL, Carter DR, Schurman DJ. Pressure and Shear Differentially Alter Human Articular Chondrocyte Metabolism: A Review. *Clin Orthop Relat Res.* Published online 2004;S89-95.

22. Li G, Yin J, Gao J, et al. Subchondral Bone in Osteoarthritis: Insight into Risk Factors and Microstructural Changes. *Arthritis Res Ther*. 2013;15:223. doi:[10.1186/ar4405](https://doi.org/10.1186/ar4405)
23. Mahjoub M, Berenbaum F, Houard X. Why Subchondral Bone in Osteoarthritis? The Importance of the Cartilage Bone Interface in Osteoarthritis. *Osteoporos Int*. 2012;23 Suppl 8:S841-846. doi:[10.1007/s00198-012-2161-0](https://doi.org/10.1007/s00198-012-2161-0)
24. Sharma AR, Jagga S, Lee SS, Nam JS. Interplay between Cartilage and Subchondral Bone Contributing to Pathogenesis of Osteoarthritis. *Int J Mol Sci*. 2013;14:19805-19830. doi:[10.3390/ijms141019805](https://doi.org/10.3390/ijms141019805)
25. Henrotin Y, Pesesse L, Sanchez C. Subchondral Bone and Osteoarthritis: Biological and Cellular Aspects. *Osteoporos Int*. 2012;23 Suppl 8:S847-851. doi:[10.1007/s00198-012-2162-z](https://doi.org/10.1007/s00198-012-2162-z)
26. Lajeunesse D, Reboul P. Subchondral Bone in Osteoarthritis: A Biologic Link with Articular Cartilage Leading to Abnormal Remodeling. *Curr Opin Rheumatol*. 2003;15:628-633. doi:[10.1097/00002281-200309000-00018](https://doi.org/10.1097/00002281-200309000-00018)
27. Benito MJ, Veale DJ, FitzGerald O, van den Berg WB, Bresnihan B. Synovial Tissue Inflammation in Early and Late Osteoarthritis. *Ann Rheum Dis*. 2005;64:1263-1267. doi:[10.1136/ard.2004.025270](https://doi.org/10.1136/ard.2004.025270)
28. Beck M, Kalhor M, Leunig M, Ganz R. Hip Morphology Influences the Pattern of Damage to the Acetabular Cartilage: Femoroacetabular Impingement as a Cause of Early Osteoarthritis of the Hip. *J Bone Joint Surg Br*. 2005;87:1012-1018. doi:[10.1302/0301-620X.87B7.15203](https://doi.org/10.1302/0301-620X.87B7.15203)
29. Bittersohl B, Steppacher S, Haamberg T, et al. Cartilage Damage in Femoroacetabular Impingement (FAI): Preliminary Results on Comparison of Standard Diagnostic vs Delayed Gadolinium-Enhanced Magnetic Resonance Imaging of Cartilage (dGEMRIC). *Osteoarthritis Cartilage*. 2009;17:1297-1306. doi:[10.1016/j.joca.2009.04.016](https://doi.org/10.1016/j.joca.2009.04.016)
30. Sandell LJ. Etiology of Osteoarthritis: Genetics and Synovial Joint Development. *Nat Rev Rheumatol*. 2012;8:77-89. doi:[10.1038/nrrheum.2011.199](https://doi.org/10.1038/nrrheum.2011.199)
31. Casartelli NC, Maffiuletti NA, Item-Glatthorn JF, et al. Hip Muscle Weakness in Patients with Symptomatic Femoroacetabular Impingement. *Osteoarthritis Cartilage*. 2011;19:816-821. doi:[10.1016/j.joca.2011.04.001](https://doi.org/10.1016/j.joca.2011.04.001)
32. Nepple JJ, Goljan P, Briggs KK, Garvey SE, Ryan M, Philippon MJ. Hip Strength Deficits in Patients With Symptomatic Femoroacetabular Impingement and Labral Tears. *Arthroscopy*. 2015;31:2106-2111. doi:[10.1016/j.arthro.2015.04.095](https://doi.org/10.1016/j.arthro.2015.04.095)
33. McCarthy JC, Noble PC, Schuck MR, Wright J, Lee J. The Otto E. Aufranc Award: The Role of Labral Lesions to Development of Early Degenerative Hip Disease. *Clin Orthop Relat Res*. Published online 2001;25-37. doi:[10.1097/00003086-200112000-00004](https://doi.org/10.1097/00003086-200112000-00004)
34. McCarthy JC, Busconi B. The Role of Hip Arthroscopy in the Diagnosis and Treatment of Hip Disease. *Orthopedics*. 1995;18:753-756. doi:[10.3928/0147-7447-19950801-12](https://doi.org/10.3928/0147-7447-19950801-12)
35. Buckwalter JA, Roughley PJ, Rosenberg LC. Age-Related Changes in Cartilage Proteoglycans: Quantitative Electron Microscopic Studies. *Microsc Res Tech*. 1994;28:398-408. doi:[10.1002/jemt.1070280506](https://doi.org/10.1002/jemt.1070280506)
36. Martin JA, Buckwalter JA. The Role of Chondrocyte Senescence in the Pathogenesis of Osteoarthritis and in Limiting Cartilage Repair. *J Bone Joint Surg Am*. 2003;85-A Suppl 2:106-110. doi:[10.2106/00004623-200300002-00014](https://doi.org/10.2106/00004623-200300002-00014)
37. Loeser RF. Age-Related Changes in the Musculoskeletal System and the Development of Osteoarthritis. *Clin Geriatr Med*. 2010;26:371-386. doi:[10.1016/j.cger.2010.03.002](https://doi.org/10.1016/j.cger.2010.03.002)
38. Vignon E, Arlot M, Patricot LM, Vignon G. The Cell Density of Human Femoral Head Cartilage. *Clin Orthop Relat Res*. Published online 1976:303-308. doi:[10.1097/00003086-197611000-00044](https://doi.org/10.1097/00003086-197611000-00044)
39. Zampogna B, Ferrini A, Zampoli A, et al. Total Hip Arthroplasty in Patients under 35 Years: A Systematic Review of the Last 2 Decades Studies. *Hip Int*. 2025;35:92-101. doi:[10.1177/11207000241289589](https://doi.org/10.1177/11207000241289589)
40. Harris EC, Coggon D. HIP Osteoarthritis and Work. *Best Pract Res Clin Rheumatol*. 2015;29:462-482. doi:[10.1016/j.berh.2015.04.015](https://doi.org/10.1016/j.berh.2015.04.015)
41. Sulsky SI, Carlton L, Bochmann F, et al. Epidemiological Evidence for Work Load as a Risk Factor for Osteoarthritis of the Hip: A Systematic Review. *PLoS One*. 2012;7:e31521. doi:[10.1371/journal.pone.0031521](https://doi.org/10.1371/journal.pone.0031521)
42. MacGregor AJ, Antoniadou L, Matson M, Andrew T, Spector TD. The Genetic Contribution to Radiographic Hip Osteoarthritis in Women: Results of a Classic Twin Study. *Arthritis Rheum*. 2000;43(11):2410-2416. doi:[10.1002/1529-0131\(200011\)43:11](https://doi.org/10.1002/1529-0131(200011)43:11)

43. Nevitt MC, Xu L, Zhang Y, et al. Very Low Prevalence of Hip Osteoarthritis among Chinese Elderly in Beijing, China, Compared with Whites in the United States: The Beijing Osteoarthritis Study. *Arthritis Rheum.* 2002;46:1773-1779. doi:[10.1002/art.10332](https://doi.org/10.1002/art.10332)
44. Jiang L, Rong J, Wang Y, et al. The Relationship between Body Mass Index and Hip Osteoarthritis: A Systematic Review and Meta-Analysis. *Joint Bone Spine.* 2011;78:150-155. doi:[10.1016/j.jbspin.2010.04.011](https://doi.org/10.1016/j.jbspin.2010.04.011)
45. Vuolteenaho K, Koskinen A, Kukkonen M, et al. Leptin Enhances Synthesis of Proinflammatory Mediators in Human Osteoarthritic Cartilage--Mediator Role of NO in Leptin-Induced PGE2, IL-6, and IL-8 Production. *Mediators Inflamm.* 2009;2009:345838. doi:[10.1155/2009/345838](https://doi.org/10.1155/2009/345838)
46. Carman WJ, Sowers M, Hawthorne VM, Weissfeld LA. Obesity as a Risk Factor for Osteoarthritis of the Hand and Wrist: A Prospective Study. *Am J Epidemiol.* 1994;139:119-129. doi:[10.1093/oxfordjournals.aje.a116974](https://doi.org/10.1093/oxfordjournals.aje.a116974)
47. Zheng L, Zhang Z, Sheng P, Mobasheri A. The Role of Metabolism in Chondrocyte Dysfunction and the Progression of Osteoarthritis. *Ageing Research Reviews.* 2021;66:101249. doi:[10.1016/j.arr.2020.101249](https://doi.org/10.1016/j.arr.2020.101249)
48. Knights AJ, Redding SJ, Maerz T. Inflammation in Osteoarthritis: The Latest Progress and Ongoing Challenges. *Current Opinion in Rheumatology.* 2023;35:128-134. doi:[10.1097/BOR.0000000000000923](https://doi.org/10.1097/BOR.0000000000000923)
49. Yi Q, Deng Z, Yue J, et al. RNA Binding Proteins in Osteoarthritis. *Front Cell Dev Biol.* 2022;10:954376. doi:[10.3389/fcell.2022.954376](https://doi.org/10.3389/fcell.2022.954376)
50. Withrow J, Murphy C, Liu Y, Hunter M, Fulzele S, Hamrick MW. Extracellular Vesicles in the Pathogenesis of Rheumatoid Arthritis and Osteoarthritis. *Arthritis Res Ther.* 2016;18:286. doi:[10.1186/s13075-016-1178-8](https://doi.org/10.1186/s13075-016-1178-8)
51. Hall M, Van Der Esch M, Hinman RS, et al. How Does Hip Osteoarthritis Differ from Knee Osteoarthritis? *Osteoarthritis and Cartilage.* 2022;30:32-41. doi:[10.1016/j.joca.2021.09.010](https://doi.org/10.1016/j.joca.2021.09.010)
52. Sun C, Zhou X, Guo T, Meng J. The Immune Role of the Intestinal Microbiome in Knee Osteoarthritis: A Review of the Possible Mechanisms and Therapies. *Front Immunol.* 2023;14:1168818. doi:[10.3389/fimmu.2023.1168818](https://doi.org/10.3389/fimmu.2023.1168818)
53. Chisari E, Wouthuyzen-Bakker M, Friedrich AW, Parvizi J. The Relation between the Gut Microbiome and Osteoarthritis: A Systematic Review of Literature. *PLoS One.* 2021;16:e0261353. doi:[10.1371/journal.pone.0261353](https://doi.org/10.1371/journal.pone.0261353)
54. Collins KH, Paul HA, Reimer RA, Seerattan RA, Hart DA, Herzog W. Relationship between Inflammation, the Gut Microbiota, and Metabolic Osteoarthritis Development: Studies in a Rat Model. *Osteoarthritis Cartilage.* 2015;23:1989-1998. doi:[10.1016/j.joca.2015.03.014](https://doi.org/10.1016/j.joca.2015.03.014)
55. Morrison DJ, Mackay WG, Edwards CA, Preston T, Dodson B, Weaver LT. Butyrate Production from Oligofructose Fermentation by the Human Faecal Flora: What Is the Contribution of Extracellular Acetate and Lactate? *Br J Nutr.* 2006;96:570-577.
56. Morrison DJ, Preston T. Formation of Short Chain Fatty Acids by the Gut Microbiota and Their Impact on Human Metabolism. *Gut Microbes.* 2016;7:189-200. doi:[10.1080/19490976.2015.1134082](https://doi.org/10.1080/19490976.2015.1134082)
57. Rastogi S, Singh A. Gut Microbiome and Human Health: Exploring How the Probiotic Genus *Lactobacillus* Modulate Immune Responses. *Front Pharmacol.* 2022;13:1042189. doi:[10.3389/fphar.2022.1042189](https://doi.org/10.3389/fphar.2022.1042189)
58. Reichardt N, Duncan SH, Young P, et al. Phylogenetic Distribution of Three Pathways for Propionate Production within the Human Gut Microbiota. *The ISME Journal.* 2014;8:1323-1335. doi:[10.1038/ismej.2014.14](https://doi.org/10.1038/ismej.2014.14)
59. Dong J, Shu G, Yang J, et al. Mechanistic Study on the Alleviation of Postmenopausal Osteoporosis by *Lactobacillus Acidophilus* through Butyrate-Mediated Inhibition of Osteoclast Activity. *Sci Rep.* 2024;14:7042. doi:[10.1038/s41598-024-57122-x](https://doi.org/10.1038/s41598-024-57122-x)
60. Feng B, Lu J, Han Y, Han Y, Qiu X, Zeng Z. The Role of Short-Chain Fatty Acids in the Regulation of Osteoporosis: New Perspectives from Gut Microbiota to Bone Health: A Review. *Medicine.* 2024;103:e39471. doi:[10.1097/MD.00000000000039471](https://doi.org/10.1097/MD.00000000000039471)
61. Lucas S, Omata Y, Hofmann J, et al. Short-Chain Fatty Acids Regulate Systemic Bone Mass and Protect from Pathological Bone Loss. *Nat Commun.* 2018;9:55. doi:[10.1038/s41467-017-02490-4](https://doi.org/10.1038/s41467-017-02490-4)
62. Tyagi AM, Yu M, Darby TM, et al. The Microbial Metabolite Butyrate Stimulates Bone Formation via T Regulatory Cell-Mediated Regulation of WNT10B Expression. *Immunity.* 2018;49:1116-1131.e7. doi:[10.1016/j.immuni.2018.10.013](https://doi.org/10.1016/j.immuni.2018.10.013)

63. Ling CW, Miao Z, Xiao ML, et al. The Association of Gut Microbiota With Osteoporosis Is Mediated by Amino Acid Metabolism: Multiomics in a Large Cohort. *J Clin Endocrinol Metab*. 2021;106:e3852-e3864. doi:[10.1210/clinem/dgab492](https://doi.org/10.1210/clinem/dgab492)
64. Zaiss MM, Joyce Wu HJ, Mauro D, Schett G, Ciccia F. The Gut–Joint Axis in Rheumatoid Arthritis. *Nat Rev Rheumatol*. 2021;17:224–237. doi:[10.1038/s41584-021-00585-3](https://doi.org/10.1038/s41584-021-00585-3)
65. Li JY, Chassaing B, Tyagi AM, et al. Sex Steroid Deficiency–Associated Bone Loss Is Microbiota Dependent and Prevented by Probiotics. *Journal of Clinical Investigation*. 2016;126:2049–2063. doi:[10.1172/JCI86062](https://doi.org/10.1172/JCI86062)
66. Redlich K, Smolen JS. Inflammatory Bone Loss: Pathogenesis and Therapeutic Intervention. *Nat Rev Drug Discov*. 2012;11:234–250. doi:[10.1038/nrd3669](https://doi.org/10.1038/nrd3669)
67. Lorenzo J. From the Gut to Bone: Connecting the Gut Microbiota with Th17 T Lymphocytes and Postmenopausal Osteoporosis. *J Clin Invest*. 2021;131:e146619. doi:[10.1172/JCI146619](https://doi.org/10.1172/JCI146619)
68. He S, Chen H, Xi H, Sun G, Du B, Liu X. Gut Microbiota-Derived Butyric Acid Alleviates Glucocorticoid-Associated Osteonecrosis of the Femoral Head via Modulating Inflammatory Cytokines in Bone Marrow Mesenchymal Stem Cells. *Mediators of Inflammation*. 2025;2025:8742817. doi:[10.1155/mi/8742817](https://doi.org/10.1155/mi/8742817)
69. Guido G, Ausenda G, Iascone V, Chisari E. Gut Permeability and Osteoarthritis, towards a Mechanistic Understanding of the Pathogenesis: A Systematic Review. *Ann Med*. 2021;53:2380–2390. doi:[10.1080/07853890.2021.2014557](https://doi.org/10.1080/07853890.2021.2014557)
70. Wei Z, Li F, Pi G. Association Between Gut Microbiota and Osteoarthritis: A Review of Evidence for Potential Mechanisms and Therapeutics. *Front Cell Infect Microbiol*. 2022;12:812596. doi:[10.3389/fcimb.2022.812596](https://doi.org/10.3389/fcimb.2022.812596)
71. Clarke TB, Davis KM, Lysenko ES, Zhou AY, Yu Y, Weiser JN. Recognition of Peptidoglycan from the Microbiota by Nod1 Enhances Systemic Innate Immunity. *Nat Med*. 2010;16:228–231. doi:[10.1038/nm.2087](https://doi.org/10.1038/nm.2087)
72. Huang ZY, Stabler T, Pei FX, Kraus VB. Both Systemic and Local Lipopolysaccharide (LPS) Burden Are Associated with Knee OA Severity and Inflammation. *Osteoarthritis Cartilage*. 2016;24:1769–1775. doi:[10.1016/j.joca.2016.05.008](https://doi.org/10.1016/j.joca.2016.05.008)
73. Hernandez CJ. The Microbiome and Bone and Joint Disease. *Curr Rheumatol Rep*. 2017;19:77. doi:[10.1007/s11926-017-0705-1](https://doi.org/10.1007/s11926-017-0705-1)
74. McAllister MJ, Chemaly M, Eakin AJ, Gibson DS, McGilligan VE. NLRP3 as a Potentially Novel Biomarker for the Management of Osteoarthritis. *Osteoarthritis Cartilage*. 2018;26:612–619. doi:[10.1016/j.joca.2018.02.901](https://doi.org/10.1016/j.joca.2018.02.901)
75. Zhao LR, Xing RL, Wang PM, et al. NLRP1 and NLRP3 Inflammasomes Mediate LPS/ATP-induced Pyroptosis in Knee Osteoarthritis. *Mol Med Rep*. 2018;17:5463–5469. doi:[10.3892/mmr.2018.8520](https://doi.org/10.3892/mmr.2018.8520)
76. Dunn CM, Velasco C, Rivas A, et al. Identification of Cartilage Microbial DNA Signatures and Associations With Knee and Hip Osteoarthritis. *Arthritis & Rheumatology*. 2020;72:1111–1122. doi:[10.1002/art.41210](https://doi.org/10.1002/art.41210)
77. Bonanzinga T, Conte P, Anzillotti G, et al. Native Intra-Articular Knee Microbiome Is a Matter of Facts: A Systematic Review of Clinical Evidence. *EFORT Open Reviews*. 2024;9:969–979. doi:[10.1530/EOR-23-0191](https://doi.org/10.1530/EOR-23-0191)
78. Berthelot JM, Sellam J, Maugars Y, Berenbaum F. Cartilage–Gut–Microbiome Axis: A New Paradigm for Novel Therapeutic Opportunities in Osteoarthritis. *RMD Open*. 2019;5:e001037. doi:[10.1136/rmdopen-2019-001037](https://doi.org/10.1136/rmdopen-2019-001037)
79. Bockenstedt LK, Gonzalez DG, Haberman AM, Belperron AA. Spirochete Antigens Persist near Cartilage after Murine Lyme Borreliosis Therapy. *J Clin Invest*. 2012;122:2652–2660. doi:[10.1172/JCI58813](https://doi.org/10.1172/JCI58813)
80. Behera AK, Durand E, Cugini C, et al. Borrelia Burgdorferi BBB07 Interaction with Integrin Alpha3beta1 Stimulates Production of Pro-Inflammatory Mediators in Primary Human Chondrocytes. *Cell Microbiol*. 2008;10:320–331. doi:[10.1111/j.1462-5822.2007.01043.x](https://doi.org/10.1111/j.1462-5822.2007.01043.x)
81. Cuevas-Sierra A, Ramos-Lopez O, Riezu-Boj JI, Milagro FI, Martinez JA. Diet, Gut Microbiota, and Obesity: Links with Host Genetics and Epigenetics and Potential Applications. *Adv Nutr*. 2019;10:S17–S30. doi:[10.1093/advances/nmy078](https://doi.org/10.1093/advances/nmy078)
82. Kattner AA. Aging like Fine Wine: Mischievous Microbes and Other Factors Influencing Senescence. *Biomedical Journal*. 2024;47:100722. doi:[10.1016/j.bj.2024.100722](https://doi.org/10.1016/j.bj.2024.100722)
83. Ma J, Zhang J, Kuang Z. A Microbiota–Epigenetic Circuit Controls Systematic Circadian Programs in the Gut Epithelium. *Front Syst Biol*. 2023;3:1175306. doi:[10.3389/fsysb.2023.1175306](https://doi.org/10.3389/fsysb.2023.1175306)

84. Zhao E, Tait C, Minacapelli CD, Catalano C, Rustgi VK. Circadian Rhythms, the Gut Microbiome, and Metabolic Disorders. *Gastro Hep Advances*. 2022;1:93-105. doi:[10.1016/j.gastha.2021.10.008](https://doi.org/10.1016/j.gastha.2021.10.008)
85. Schott EM, Farnsworth CW, Grier A, et al. Targeting the Gut Microbiome to Treat the Osteoarthritis of Obesity. *JCI Insight*. 2018;3:e95997. doi:[10.1172/jci.insight.95997](https://doi.org/10.1172/jci.insight.95997)
86. Huang MH, Liu CX, Huang YS, He TY, Dong ZQ. Genetic Association of Gut Microbiota with Osteoarthritis: A Multivariable Mendelian Randomization Study Considering Medication Use. *J Med Microbiol*. 2024;73. doi:[10.1099/jmm.0.001920](https://doi.org/10.1099/jmm.0.001920)
87. Cintio M, Scarsella E, Sgorlon S, Sandri M, Stefanon B. Gut Microbiome of Healthy and Arthritic Dogs. *Veterinary Sciences*. 2020;7:92. doi:[10.3390/vetsci7030092](https://doi.org/10.3390/vetsci7030092)
88. Li J, Liang J, Liu Y, Sun W, Sun W. Basal Metabolic Rate Mediates the Causal Relationship between Gut Microbiota and Osteoarthritis: A Two-Step Bidirectional Mendelian Randomization Study. *Front Microbiol*. 2024;15:1371679. doi:[10.3389/fmicb.2024.1371679](https://doi.org/10.3389/fmicb.2024.1371679)
89. Lee YH, Song GG. The Gut Microbiome and Osteoarthritis: A Two-Sample Mendelian Randomization Study. *J Rheum Dis*. 2021;28:94-100. doi:[10.4078/jrd.2021.28.2.94](https://doi.org/10.4078/jrd.2021.28.2.94)
90. Goswami K, Clarkson S, Tipton C, et al. The Microbiome of Osteoarthritic Hip and Knee Joints: A Prospective Multicenter Investigation. *J Bone Joint Surg Am*. 2023;105:821-829. doi:[10.2106/JBJS.22.00594](https://doi.org/10.2106/JBJS.22.00594)
91. Fernández-Rodríguez D, Baker CM, Tarabichi S, Johnson EE, Ciccotti MG, Parvizi J. Mark Coventry Award: Human Knee Has a Distinct Microbiome: Implications for Periprosthetic Joint Infection. *J Arthroplasty*. 2023;38:S2-S6. doi:[10.1016/j.arth.2023.03.084](https://doi.org/10.1016/j.arth.2023.03.084)
92. Anderson PM, Frank T, Herz M, et al. Qualitative Comparison of Cultured Skin Microbiota From the Inguinal Region of Obese and Nonobese Patients Eligible for Hip Arthroplasty. *Arthroplasty Today*. 2024;30:101483. doi:[10.1016/j.artd.2024.101483](https://doi.org/10.1016/j.artd.2024.101483)
93. Karim A, Khan HA, Iqbal MS, Ahmad F, Qaisar R. Probiotics' Supplementation Alleviates Disease Severity and Improves Postural Balance by Repairing Intestinal Leak in Patients Suffering from Osteoarthritis: A Double-Blinded Clinical Trial. *Br J Nutr*. 2024;132:1602-1610. doi:[10.1017/S0007114524002824](https://doi.org/10.1017/S0007114524002824)
94. Lei M, Guo C, Wang D, Zhang C, Hua L. The Effect of Probiotic *Lactobacillus Casei Shirota* on Knee Osteoarthritis: A Randomised Double-Blind, Placebo-Controlled Clinical Trial. *Benef Microbes*. 2017;8:697-703. doi:[10.3920/BM2016.0207](https://doi.org/10.3920/BM2016.0207)
95. Pedersini P, Savoldi M, Berjano P, Hugo Villafañe J. A Probiotic Intervention on Pain Hypersensitivity and Microbiota Composition in Patients with Osteoarthritis Pain: Study Protocol for a Randomized Controlled Trial. *Arch Rheumatol*. 2021;36:296-301. doi:[10.46497/ArchRheumatol.2021.7719](https://doi.org/10.46497/ArchRheumatol.2021.7719)
96. Amabebe E, Robert FO, Agbalalah T, Orubu ESF. Microbial Dysbiosis-Induced Obesity: Role of Gut Microbiota in Homeostasis of Energy Metabolism. *Br J Nutr*. 2020;123:1127-1137. doi:[10.1017/S0007114520000380](https://doi.org/10.1017/S0007114520000380)